METHODS AND REAGENTS FOR THE DETECTION OF MELANOMA BACKGROUND OF THE INVENTION

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Cutaneous malignant melanoma is a common, aggressive cancer with growing incidence. It is a serious healthcare problem with over 55,100 new cases anticipated in 2004 in US, and a mortality rate of about 14.5%. Cancer Facts and Figures 2003. American Cancer Society, 2003. The incidence of melanoma continues to rise faster than that of any other malignancy. De Braud et al. (2003). While prognosis of early local melanoma is favorable with 5-year overall survival over 90%, regional lymph node involvement decreases the overall survival rate to 10-46%. Balch et al. (2001). Therefore regional lymph node (LN) status becomes the most significant prognostic factor in a melanoma patient's survival. Introduction of the sentinel lymph nodes (SLN) technique (Morton (1992)) has increased the sensitivity of melanoma micrometastasis detection compared to H&E staining alone. Yu et al. (1999); and Messina et al. (1999). Nevertheless, even when enhanced by IHC, histological analysis is limited by the ability of light microscopy to recognize the tumor cells. Reverse transcription-polymerase chain reaction (RT-PCR) analysis has recently been proposed for a more sensitive detection of melanoma cells in LN. Many studies, when using well-characterized melanocyte specific markers, such as tyrosinase and MART-I, has/e demonstrated the presence of these gene transcripts in LNs oi.ierwise found to be negative by routine histology and IHC. Shivers et al. (1998); and Kuo et al. (2003). However, these genes are not specific to tumor cells and cannot be used to discriminate between benign and malignant tissue. hi fact, they caused false-positive results in the presence of benign capsular nevi. Takeuchi et al. (2004); Starz et al. (2003); and Gutzmer et al. (2002). Considering that benign nevi are not rare events in the melanoma SLN, the current RT-PCR assays are not useful clinically for diagnostic of melanoma micrometastasis. A recent study, proposed a multi-marker panel, including cancer specific markers for RT-PCR assay in order to increase assay specificity. Hoon et al. (2004). Identification of novel melanoma specific markers remains one of the key questions of melanoma research.

Certain proteins have been shown to be associated with melanoma and its metastases. These proteins or their activities have been used in IHC to identify

metastases and include LlCAM (Thies et al. (2002); Fogel et al. (2003)); and S-100 (Diego et al. (2003)).

Nucleic acid tests have been proposed to increase the sensitivity of detection of metastatic melanoma. US Patent Publication Nos. 2002/01 10820; and 2003/0232356. Studies have used markers that include MAGE3, tyrosinase, MART-1, MITF-M or 5 IL-I, Rl, endothelin-2, ephrin-A5, IGF Binding protein 7, HLA-A0202 heavy chain, Activin A (βA subunit), TNF RII, SPC4, CNTF Ra, or gplOO (HMB45) genes. Bostick et al. (1999); Hoon et al. (2001); Palmieri et al. (2001); Wrightson et al. (2001); Gutzmer et al. (2002); Davids et al. (2003); Starz et al. (2003); Rimboldi et al. (2003); Cook et al (2003); Reintgen et al. (2004); US Patent Publication Nos. 10 2002/0098535; 2003/0049701; US Patent Nos. 5,512,437; 5,512,444; 5,612,201; 5,759,783; 5,844,075; 6,025,474; 6,057,105; 6,235,525; 6,291,430; 6,338,947; 6,369,21 1; 6,426,217; 6,475,727; 6,500,919; 6,527,560; 6,599,699; WO 96/29430. Where determined, these markers have not been found adequate for sole use in 15 melanoma diagnosis. Riccioni et al. (2002); Gutzmer et al. (2002); Davids et al. (2003); Goydos et al. (2003); and Prichard et al. (2003).

A number of these markers have also been shown to be indicative of other neoplasias such as ME20M (GPlOO) for clear cell sarcoma, biliary tract carcinoma and gastric carcinoma. Hiraga et al. (1997); Okada et al. (2001); Okami et al. (2001); Antonescu et al. (2002); Segal et al. (2003). MAGE3 is also indicative of a number of neoplasias including breast, hepatocellular, renal, neural, lung and esophageal. Yamanaka et al. (1999); Ooka et al. (2000); Suzuki et al. (2000); Cheung et al. (2001); and Weiser et al. (2001). Several melanoma antigen-encoding genes are also expressed in lung cancer. Yoshimatsu et al. (1998).

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These markers proved to be sensitive but non-specific since they showed positive expression in other cancers and benign melanocytes. Additionally, tyrosinase is expressed in Schwann cells which are present in normal lymph nodes. The lack of specificity alone calls for the development of assays with new or additional markers. H&E histology and IHC remain the "gold standard for the identification of melanoma and nevus cells in SLNs." Starz et al. (2003). Detection issues in the intra-operative setting make this need even more acute.

Lymph node involvement is the strongest prognostic factor in many solid tumors, and detection of lymph node micrometastases is of great interest to pathologists and surgeons. Current lymph node evaluation involves microscopic examination of H&E-stained tissue sections and IHC and suffers from three major limitations: (a) small foci of cells, are easily missed; (b) the result is not rapidly available, meaning that any positive result in a SLN procedure requires a second surgery for removal of axcillary lymph nodes; and (c) only one or two tissue sections are studied, and thus the vast majority of each node is left unexamined. Serial sectioning can help overcome sampling error, and IHC can help identify small foci of cells; this combination, however, is costly and time consuming for routine analysis.

Surgical decisions of regional lymph node dissection can be based on intra-operative frozen section analysis of lymph nodes; however, the sensitivity of these methods is relatively poor, ranging from 50-70% relative to standard H&E pathology, leading to a high rate of second surgeries. Thus, pathologists are not routinely performing intra-operative frozen section analysis or touch print cytology analysis for melanoma patients. Improvements in the sensitivity and specificity of intra-operative assays for melanoma would significantly benefit oncology.

High-density microarrays have been applied to simultaneously monitor expression, in biological samples, of thousands of genes. Studies have resulted in the identification of genes differentially expressed in benign and malignant lesions, as well as genes that might be of prognostic value. Luo et al. (2001); and Wang et al. (2004). Gene expression profiling of malignant melanoma has been accomplished using a microarray containing probes for 8,150 cDNAs. Bittner et al. (2000). These researchers identified several genes that might be associated with aggressive tumor behavior. In recent work, comparison of gene expression profiles of a few melanoma and normal melanocyte cell lines led to the identification of differentially expressed genes and pathways modulated in melanoma. Takeuchi et al. (2004).

SUMMARY OF THE INVENTION

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Gene expression profiling of an extensive set of clinically relevant tissue samples is provided in the present invention. Total RNA from forty-five primary malignant melanomas, 18 benign skin nevi and 7 normal skin tissues were hybridized on an

Affymetrix HuI 33A microarray containing 22,000 probe sets. Differentially expressed genes in malignant melanoma as compared to benign tissue were identified. Pathway analysis of the differentially expressed genes revealed an over-representation of genes associated with neural tissue development and activation of amyloid processing signaling pathway. A one-step quantitative RT-PCR assay was used to test a combination of two melanoma specific genes, PLAB and LlCAM in a panel of clinically relevant samples that included primary malignant melanoma, benign nevi, melanoma LN metastasis and melanoma-free lymph node samples.

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The present invention provides a method of identifying a melanoma by obtaining a tissue sample; and assaying and measuring the expression levels in the sample of genes encoding mRNA corresponding to prostate differentiation factor (PLAB, MICI) (SEQ ID NO: 1) and L1 cell adhesion molecule (LICAM) (SEQ ID NO: 2); or PLAB, LICAM and neurotrophic tyrosine kinase receptor, type 3 (NTRK3) (SEQ ID NO: 3) where the gene expression levels above pre-determined cut-off levels are indicative of the presence of a melanoma in the sample. The invention further provides a method of identifying a melanoma by obtaining a tissue sample; and assaying and measuring the expression levels in the sample of genes encoding mRNA recognized by the primer/probe sets SEQ ID NOs: 4-6 or SEQ ID NOs: 7-9 and SEQ ID NOs: 10-12 or SEQ ID NOs: 13-15; or SEQ ID NOs: 4-6 or SEQ ID NOs: 7-9 and SEQ ID NOs: 10-12 or SEQ ID NOs: 13-15 and SEQ ID NOs: 16-18 where the gene expression levels above pre-determined cut-off levels are indicative of the presence of a melanoma in the sample.

The invention also provides a method of distinguishing a malignant melanocyte from a benign melanocyte by obtaining a tissue sample; and assaying and measuring the expression levels in the sample of genes encoding PLAB and LICAM; or PLAB, LICAM and NTRK3 where the gene expression levels above pre-determined cut-off levels are indicative of the presence of a melanoma in the sample.

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SEQ ID NOs: 4-6 or SEQ ID NOs: 7-9 and SEQ ID NOs: 10-12 or SEQ ID NOs: 13-15 and SEQ ID NOs: 16-18 where the gene expression levels above pre-determined cut-off levels are indicative of the presence of a melanoma in the sample.

The invention further provides a method of determining patient treatment protocol by obtaining a tissue sample from the patient; and assaying and measuring the expression levels in the sample of genes encoding PLAB and LlCAM; or PLAB, LlCAM and NTRK3 where the gene expression levels above pre-determined cut-off levels are indicative of the presence of a melanoma in the sample.

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The invention further provides additional Marker and control genes, the expression of which aid in the claimed methods. These additional genes include up-regulated SEQ ID NOs: 29-467 and down-regulated SEQ ID NOs: 468-978.

The primary Marker can be PLAB and is defined herein as the gene encoding any variant, allele etc. including SEQ ID NO: 1. PLAB is also described by Paralkar et al. (1998) and represented by Accession No. AF003934. PLAB is also defined as the gene encoding mRNA recognized by the primer/probe sets SEQ ID NOs: 4-9.

The secondary Marker can be LICAM and is defined herein as the gene encoding any variant, allele etc. including SEQ ID NO: 2. LICAM is also described by Haspel et al (2003); and US Patent No. 6,107,476 and is represented by Accession No. NM_000425. LICAM is also defined as the gene encoding mRNA recognized by the primer/probe sets SEQ ID NOs: 10-15.

The invention further provides a kit for conducting an assay to determine the presence of melanoma in a cell sample comprising: nucleic acid amplification and detection reagents.

The invention further provides primer/probe sets for amplification and detection of PCR products obtained in the inventive methods. These sets include the following:

- SEQ ID NO:4 (PLAB forward primer) ggcagaatcttcgtccgca
- SEQ ID NO: 5(PLAB reverse primer) ggacagtggtccccgttg
- 5 SEQ ID NO.6 (PLAB probe) cccagctggagttgcacttgcggcc
 - SEQ ID NO:7 (PLAB upper primer) gaacaccgacctcgtccc
 - SEQ ID NO: 8 (PLAB lower primer) ggcggcccgagagata
 - SEQ ID NO: 9 (PLAB probe) cgccagaagtgcggctgggattt
 - SEQ ID NO: 10 (LICAM forward) gctgggactgggaacagaact
- 10 SEQ ID NO:11 (L1CAM Reverse) ggagcagagatggcaaagaaa
 - SEQ ID NO:12 (L1CAM probe) ttccccaccatctgctgt
 - SEQ ID NO: 13 (LlCAM upper) ccacagatgacatcagcctcaa
 - SEQ ID NO: 14 (L1CAM lower) ggtcacacccagctcttcctt
 - SEQ ID NO:15 (LlCAM probe) tggcaagcccgaagtgcagttcctt
- 15 SEQ ID NO:16 (NTRK3 primer) gccccggcacccttta
 - SEQ ID NO:17 (NTRK3 primer) aaccetgccagtggtggat
 - SEQ ID NO: 18 (NTRK3 probe) cagatgggtgttttc
 - SEQ ID NO: 19 (Tyr upper) acteagcccagcatcattette
 - SEQ ID NO:20 (Tyr lower) atggctgttgtactcctccaatc
- 20 SEQ ID NO:21 (Tyr probe) cttctcctcttggcagattgtctgtagctt
 - SEQ ID NO:22 (PBGD upper) ccacacacagcctactttccaa
 - SEQ ID NO:23 (PBGD lower) tacccacgcgaatcactctca
 - SEQ ID NO:24 (PBGD probe) aacggcaatgcggctgcaacggcggaatt
 - The invention further provides amplicons obtained by PCR methods utilized in the
- 25 inventive methods. These amplicons include the following:
 - SEQ ID NO:25 (PLAB Amplicon)
 - gaacaccgacctcgtcccggcccctgcagtccggatactcacgccagaagtgcggctgggatccggcggccacctgcacctgcatctctccgggccgcc
 - SEQ ID NO:26 (LlCAM Amplicon)
- ccacagatgacatcagcctcaagtgtgaggccagtggcaagcccgaagtgcagttccgctggacgagggatggtgtccacttcaaacccaaggaagagctgggtgtgacc
 - SEQ ID NO:27 (tyrosinase Amplicon)
 - actcagcccagcatcattcttctcctcttggcagattgtctgtagccgattggaggagtacaacagccat

SEQ ID NO:28 (PBGD Amplicon)

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 ${\tt ccacacacagcctactttccaagcggagccatgtctggtaacggcaatgcggctgcaacggcggaagaaaacagcccaaagatgagagtgattcgcgtgggta}$

Other genes described herein include up-regulated Markers (SEQ ID NOs: 29-467), down-regulated Markers (SEQ ID NOs: 468-978), PBGD (SEQ ID NO: 979), MARTI (SEQ ID NO: 980), ME20M (GPIOO; SEQ ID NO: 981) and MAGE-3 (SEQ ID NO: 982) and various primers and probes (SEQ ID NOs: 983-101 1) used in detecting their expression.

BRIEF DESCRIPTION OF THE DRAWINGS

- 10 Fig. 1. Flowchart of data analysis.
 - Fig. 2. Hierarchical clustering on the 15,795 genes that have at least two "present" calls in all samples. Each column is a sample and each row is a gene. Red is up-regulation and green is down-regulation. Purple: melanoma samples; yellow: benign nevi; and blue: normal skin.
- Fig. 3. Microarray expression (A) and real time RT-PCR validation data (B) of the selected genes. First fourteen samples from the left are the melanoma tissue samples (red); next seven are benign nevi samples (yellow) and last five are normal skin (blue). For microarray plots x-axis shows intensity values; for PCR plots, x—axis is 2^{ΔCT}, where ΔCt is Ct (Target Gene) Ct PBGD.
 - Fig. 4. Amyloid processing pathway. Adopted from Ingenuity™ Pathway

 Analysis Software Application. Genes up-regulated in melanoma are red and downregulated in melanoma are green. Each gene symbol is followed by the fold-change
 of expression level between melanoma and benign/normal samples.
- Fig. 5. One-step quantitative RT-PCR assay of PLAB and LlCAM (A) and conventional melanoma markers, gplOO, tyrosinase (SEQ ID NO: 999) and MARTI (B). For each plot x-axis represents score for the new markers or the conventional markers. Median scores for each samples category are labeled. Two cut-off levels based on normal (green) and benign (red) samples are labeled on each plot.

DETAILED DESCRIPTION

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The present invention provides methods of qualitatively and quantitatively identifying a melanoma; distinguishing a malignant melanocyte from a benign melanocyte; diagnosing melanocytic lesions with uncertain pathological features; and determining a melanoma patient treatment protocol. The methods further provide aids in patient prognosis, patient monitoring and drug development. The methods rely on assaying and measuring expression levels of various Marker genes encoding mRNAs provided herein where gene expression over a pre-determined cut-off level is indicative of the presence of a malignant melanocyte in the sample assayed.

Cutaneous melanoma is a common, aggressive cancer with growing incidence. Identification of melanoma-specific deregulated genes could provide molecular markers for LN staging assays and further insight on melanoma tumorigenesis. Total RNA isolated from 45 primary melanoma, 18 benign skin nevi, and 7 normal skin tissue specimens were analyzed on an Affymetrix U133A microarray containing 22,000 probe sets. Hierarchical clustering revealed a distinct separation of the melanoma samples from benign and normal specimens. Novel genes associated with malignant melanoma were identified. Differential gene expression of two melanoma specific genes, PLAB and LICAM, were tested by a one-step quantitative RT-PCR assay on primary malignant melanoma, benign nevi and normal skin samples and also on malignant melanoma LN metastasis and melanoma-free lymph nodes. The performance of the markers was compared to conventional melanoma markers such as tyrosinase, gplOO, and MARTI. The results demonstrated the ability of using a combination of PLAB and LICAM in a RT-PCR assay to differentiate clinically relevant tissue samples containing benign or malignant melanocytes.

High-density cDNA and oligonucleotide microarrays allow simultaneous monitoring of the expression of thousands of genes. Microarray technology provides a quantitative measurement of mRNA abundance and has gained acceptance as a tool for marker discovery based on gene expression. In the context of cancer research, microarray analysis has identified genes differentially expressed in benign and malignant lesions, different cancer types or that have prognostic significance. Luo et al. (2001); Su et al. (2001); Henshall et al. (2003); and Wang et al. (2004). The first

gene expression profiling of malignant melanoma used a microarray containing probes for 8,150 cDNAs and identified genes that might be associated with aggressive tumor behavior. Bittner et al. (2000). Since the samples analyzed in their study did not include tissues containing normal or benign melanocytes, differentially expressed genes in malignant melanoma were not identified. hi contrast to normal skin, melanocyte content in benign nevi is close to that in melanoma.

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In another study, two pooled samples derived from either melanoma or benign nevi tissues were hybridized to a cDNA array and genes preferentially expressed in melanoma- or nevi-derived samples were found. Seykora et al. (2003). Other researchers used subtractive hybridization or analysis of SAGE libraries generated on melanoma cell lines, for monitoring gene expression in melanoma. Hipfel et al. (2000); and Weeraratna (2004). Recently, comparison of gene expression profiles of a few melanoma and melanocyte cell lines led to the identification of differentially expressed genes and pathways modulated in melanoma. Hock et al. (2004). While these studies provide a solid foundation for melanoma genetics, there is no marker that can clearly differentiate melanoma from benign tissue. Several markers currently used such as tyrosinase and Mart-1 cannot discriminate between benign and malignant tissue. Takeuchi et al. (2004). Consequently, these markers have limited use in applications such as intra-operative, lymph-node-based staging of disease.

Difficulties in obtaining sufficient RNA samples from malignant melanoma and benign melanocyte lesions, tissue heterogeneity, and the presence of melanin in purified RNA remain the major challenges in these studies. In the study presented herein, total RNA isolated from 45 primary malignant melanomas, 18 benign skin nevi, and 7 normal skin tissues were hybridized on an Affymetrix HuI 33A microarray containing 22,000 probe sets. A modified RNA extraction method was developed to produce melanin-free RNA samples that increased the micorarray hybridization signals. Hierarchical clustering revealed distinct separation of the melanoma samples from benign and normal specimens. Significance Analysis of Microarray (SAM) method, *t-test* and percentile analysis identified 439 up-regulated (SEQ ID NOs: 29-467) and 511 down-regulated (SEQ ID NOs: 468-978) genes in the melanoma samples. Besides well-characterized genes such as me20m (gplOO), melanocortin

receptor 1, and LlCAM, many novel genes previously unassociated with melanoma were identified including NTKR3 and PLAB. Pathway analysis of the differentially expressed genes revealed an over-representation of genes associated with neural tissue development and function, activation of amyloid processing and integrin signalling pathways. RT-PCR assays were performed to confirm the differential expression of the selected genes.

The methods provided have sufficient specificity and sensitivity to detect metastasis of melanoma. A comparison of the current methods available indicates that tradition methods of H&E and IHC are clinically acceptable whereas, prior to the current invention, PCR methods were unacceptable. Table 1 shows the drawbacks and advantages of current methods prior to the invention claimed herein.

Table 1

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Method	Sensitivity	Specificity
H&E	Low	100%
IHC	Low	100%
PCR	High	Low

In the present invention, specificity is preferably at least 95%, more preferably, specificity is at least 97% and most preferably, specificity is at least 99% based on a comparison of H&E and IHC negative nodes. Preferably, sensitivity is at least at least 80%, more preferably sensitivity is at least 85% and most preferably, sensitivity is at least 90% based on a comparison of H&E and IHC positive nodes. Preferably, specificity and sensitivity are at least 97% based on a comparison of H&E and IHC negative nodes and at least 85% based on a comparison of H&E and IHC positive nodes, respectively.

Preferably, the pre-determined cut-off levels are at least two-fold over-expression in tissue having metastatic melanoma relative to benign melanocyte or normal tissue.

The preferred methods of the invention employ a rapid technique for extracting nucleic acids from a tissue sample and a method of amplifying and detecting nucleic acid fragments indicative of metastasis. The nucleic acid fragments qualitatively and quantitatively measure mRNA encoded by the Marker genes. Tissue samples include lymph node, both regional and sentinel, skin lesions and other biopsy material.

The methods provided herein allow for intra-operative detection of micrometastases allowing a physician to determine whether to excise additional lymph nodes and to immediately implement an appropriate treatment protocol. As shown in Table 2, if a LN is found to be positive for melanoma, regional LNs are excised and interferon therapy could be suggested. Standard biopsy methods can take over one week and a positive result requires additional surgery to remove LNs and there is a concomitant delay in interferon therapy.

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Table 2

Clinical Stage	1° Tumor (T)	LN	Metastasis	Treatment
Stage I	T1: ≤ 1 mm	Negative	Absent	Excision 1 cm
	T2: 1.01-2.00	Negative	Absent	Excision 1-2 cm
	mm			
Stage II	T3: 2.01-4.00	Negative	Absent	Excision 2 cm
	mm			
	T4: > 4.01 mm	Negative	Absent	Excision 2 cm
Stage III	Any thickness	Positive	Absent	Excision + complete LN
				dissection + interferon
				clinical trial
Stage IV	Any thickness	Positive	Present	Interferon clinical trial,
				symptomatic therapy

It is important to adequately sample the tissue used to conduct the assay. This includes proper excision and processing of the tissue sample as well as extraction of RNA. Once obtained, it is important to process the tissue samples properly so that any cancerous cells present are detected.

In the most preferred embodiment of the invention, node sampling is also given attention both intra- and extra-operatively. Since the distribution of cancer cells in nodes is non-uniform, it is preferable that multiple sections of the node be sampled. Every identified SLN should be submitted for pathological evaluation. SLN material is ordinarily be fixed in formalin and examined as formalin fixed, paraffin embedded tissue sample. Equally representative parts of SLN are processed for molecular analysis (fresh tissue) and histology (fixed tissue). General LN sampling procedures are described in Cochran et al. (2001); and Cochran et al. (2004). One method for accomplishing both a molecular based test and an examination of the same node sample by pathology is to bisect the node through the longest diameter. Each half is then divided into at least four full-faced sections with at least one outer and inner

section for pathology as fixed material, and at least one outer and inner section for molecular testing. As the distribution of metastases and micrometastases is not uniform in nodes or other tissues, a sufficiently large sample should be obtained so that metastases will not be missed. One approach to this sampling issue in the present method is to homogenize a large tissue sample, and subsequently perform a dilution of the well-mixed homogenized sample to be used in subsequent molecular testing.

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In the case of LN tissue samples, it is preferable to remove any adipose tissue prior to cellular disruption. Manual cell and tissue disruption can be by any means known in the art such as a disposable tissue grinder described in US Patent 4,715,545 or a commercial homogenizer such as Omni GLHl 15 with disposable probes (Omni International, Warrenton, VA). Homogenization time is within 1 to 2 minutes and is more preferably 30-45 sec. The sample can then be processed to purify the RNA prior to assaying and measuring Marker expression levels. Suitable RNA purification methods include columns such as (e.g., RNeasy mini column, QIAshredder, QIAGEN Inc., Valencia, CA, or a suitable substitute).

A variety of techniques are available for extracting nucleic acids from tissue samples. Typical commercially available nucleic acid extraction kits take at least 15 minutes to extract the nucleic acid. In the preferred intra-operative methods of the instant invention, nucleic acid is extracted in less than 8 minutes and preferably less than 6 minutes.

The successful isolation of intact RNA generally involves four steps: effective disruption of cells or tissue, denaturation of nucleoprotein complexes, inactivation of endogenous ribonuclease (RNase) and removal of contaminating DNA and protein. The disruptive and protective properties of guanidinium isothiocyanate (GITC) and β-mercaptoethanol (β-me) to inactivate the ribonucleases present in cell extracts make them preferred reagents for the first step. When used in conjunction with a surfactant such as sodium dodecylsulfate (SDS), disruption of nucleoprotein complexes is achieved allowing the RNA to be released into solution and isolated free of protein. Tissues are homogenized in the GITC-containing lysis buffer, addition of ethanol creates the appropriate conditions for RNA to bind to the silica membrane. Centrifugation can clear the lysate of precipitated proteins and cellular DNA and is

preferably performed through a column. RNA purification is preferably conducted on a spin column containing silica or other material.

RNA is precipitated via the spin column as described above and centrifugation times are preferably no greater than 30 sec. Typically, the sample is diluted with an equal volume of 70% ethanol and thoroughly mixed prior to applying to the column. After washing, the column is dried by centrifugation, and RNA is eluted in RNase free water and collected by centrifugation. The total time of this rapid protocol is less than 8 minutes and preferably less than 6 min.

In summary the rapid RNA extraction method involves the following steps:

10 obtaining a tissue sample;

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homogenizing the tissue to produce a homogenate;

contacting the homogenate with a substrate containing, or to which is affixed, an RNA-binding material;

allowing the RNA to bind to the RNA binding material;

washing the substrate under conditions sufficient to remove any contaminants, interferents and un-bound RNA; and eluting bound RNA from the substrate.

The reagents involved in this rapid extraction process can be those provided by the manufacturer or can be, for instance:

Lysis/Binding buffer (preferably, 4.5M GITC, 100mM NaPO₄),
Wash buffer I (preferably, 37% ethanol in 5M GITC, 20mM Tris-HCl),
Wash buffer II (preferably, 80% ethanol in 20mM NaCl, 2mM Tris-HCl), and
Nuclease-free sterile double distilled water for elution.

In one method, prior to the process for isolating nucleic acids described above, tissue samples are weighed and put into 8 or 14 ml polypropylene culture tubes and pre-cooled on dry ice. The frozen tissue samples are then divided into pieces of about 50 mg or less without being thawed. All buffers are those provided by QIAGEN in the RNeasy mini kit. A volume of homogenization (lysis) buffer is added to the tissue based on Table 3.

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Table 3

Tissue Weight (mg)	Homogenization buffer (ml)
≤ 100	2
100-149	2
150-199	3
200-249	4
250-299	5
300-349	6
350-399	7
400-449	8
450-400	9
500-550	10
>550	*

* Tissue above 550 mg is divided into equivalent parts and processed as individual samples. An alternative method to calculate lysis buffer volume for tissues over 100 mg is to add 1 ml per 50mg tissue; using 2 ml for tissues less than 100 mg.

The tissue sample is then homogenized for instance by the Omni GLHl 15 at a power setting to grade 6, Adaptor AlOOO and disposable probes. The homogenate is then mixed with an equal volume of 70% ethanol and thoroughly mixed for instance by vortexing on a VWR Model G560 set at 10 speed (maximum) about 10 seconds or by pipetting 4-5 times. The homogenate/ethanol mixture is then applied to an RNeasy mini column mounted on a vacuum manifold in a volume in accordance with Table 4 so that a consistent amount of the original tissue (approximately 5 mg/column) is loaded thus producing comparable RNA yields for each tissue sample.

Table 4

Tissue weight	Volume homogenate/ethanol
(mg)	mix (µl) (recommended)
30-39	700
40-49	500
50-59	400
60-69	350
70-79	300
80-89	250
90-99	225
> 100	200

A vacuum is then applied to the column to remove the liquid. The vacuum is stopped and two washes of 700 ml are applied, first with RWI buffer and second with

RPE buffer each removed by filtration. Vacuum is at 800-1200 mBar in each case. The column is then placed into a 1.5 ml collection tube and centrifuged in an Eppendorf 5415D centrifuge at 13,200 rpm for 30 seconds to dry. The column is transferred to a new 1.5 ml collection tube. Fifty μ l RNase-free water is directly added to the membrane and the column is centrifuged in an Eppendorf 5415D centrifuge for 30 seconds at 13,200 rpm to elute the RNA. The RNA quality is determined with an Agilent Bioanalyzer and the RNA is stored at -70 °C.

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Melanin can negatively impact the efficiency of reversed transcription and amplification reactions. Accordingly, a melanin removal process is undertaken when the sample is suspected of containing a significant amount of melanin (as in the case of samples of a primary melanoma or benign skin nevi) and is less of a concern when performing the assay on a SLN since melanocyte content is low. If necessary, melanin is removed to enhance reverse transcription and/or nucleic acid amplification.

Typically, melanin is removed during the filtration steps provided above. In the case of tissue with high melanin concentration, less tissue should be used, approximately 5 mg per Qiagen RNeasy mini column.

If another method is used that results in residual melanin in the sample, removal involves the use of a matrix employing a polymer bead system such as Bio-Gel P-60 (Bio-Rad Laboratories, Hercules, CA). Such a method is described by Satyamoorthy et al. (2002). Essentially, this method involves preparing a 50% (w/v) mixture of the Bio-Gel material in 10 mM sodium acetate (pH 4.2). About 300 μ l of the mixture are placed in a micro-centrifuge tube and centrifuged at 1000rpm for 1 min. The supernatant is discarded and the beads are placed in a mini-column or similar vessel. Homogenate is then passed through the vessel containing the beads (after first incubating them in the vessel). The supernatant is collected. Further washing of the beads with additional 100 μ l aliquots of 10 mM sodium acetate can be used to capture additional volumes of melanin-free sample if necessary for adequate assay volume. The dark melanin will be clearly visible on the beads retained in the vessel. Other silica-based filters can also be used to remove the melanin pigment as described by Wang et al. (2001).

An important aspect of the intra-operative methods of the invention is rapid Marker detection. Provided that such methods can be conducted within a period acceptable for an intra-operative assay (i.e., no more than about 35 minutes), any reliable, sensitive, and specific method can be used.

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In the case of measuring mRNA levels to determine gene expression, assays can be by any means known in the art and include methods such as PCR, Rolling Circle Amplification (RCA), Ligase Chain Reaction (LCR), Strand Displacement Amplification (SDA), Nucleic Acid Sequence Based Amplification (NASBA), and others. The rapid molecular diagnostics involved are most preferably quantitative PCR methods, including QRT-PCR. Detection can be by any method known in the art including microarrays, gene chips and fluorescence.

A typical PCR includes multiple amplification steps, or cycles that selectively amplify target nucleic acid species. A typical PCR includes three steps: a denaturing step in which a target nucleic acid is denatured; an annealing step in which a set of PCR primers (forward and backward primers) anneal to complementary DNA strands; and an elongation step in which a thermostable DNA polymerase elongates the primers. By repeating this step multiple times, a DNA fragment is amplified to produce an amplicon, corresponding to the target DNA sequence. Typical PCR includes 20 or more cycles of denaturation, annealing and elongation. Often, the annealing and elongation steps can be performed concurrently, in which case the cycle contains only two steps.

In the preferred inventive method, employing RT-PCR, the RT-PCR amplification reaction is conducted in a time suitable for intra-operative diagnosis, the lengths of each of these steps can be in the seconds range, rather than minutes. Specifically, with certain new thermal cyclers being capable of generating a thermal ramp rate of at least about 5C° per second, RT-PCR amplifications in 30 minutes or less are used. More preferably, amplifications are conducted in less than 25 minutes. With this in mind, the following times provided for each step of the PCR cycle do not include ramp times. The denaturation step may be conducted for times of 10 seconds or less. In fact, some thermal cyclers have settings for "0 seconds" which may be the optimal duration of the denaturation step. That is, it is enough that the thermal cycler reaches

the denaturation temperature. The annealing and elongation steps are most preferably less than 10 seconds each, and when conducted at the same temperature, the combination annealing/elongation step may be less than 10 seconds. Some homogeneous probe detection methods, may require a separate step for elongation to maximize rapid assay performance. In order to minimize both the total amplification time and the formation of non-specific side reactions, annealing temperatures are typically above 50°C. More preferably annealing temperatures are above 55°C.

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A single combined reaction for RT-PCR, with no experimenter intervention, is desirable for several reasons: (1) decreased risk of experimenter error; (2) decreased risk of target or product contamination; and (3) increased assay speed. The reaction can consist of either one or two polymerases. In the case of two polymerases, one of these enzymes is typically an RNA-based DNA polymerase (reverse transcriptase) and one is a thermostable DNA-based DNA polymerase. To maximize assay performance, it is preferable to employ a form of "hot start" technology for both of these enzymatic functions. US Patents 5,41 1,876 and 5,985,619 provide examples of different "hot start" approaches. Preferred methods include the use of one or more thermoactivation methods that sequester one or more of the components required for efficient DNA polymerization. US Patents 5,550,044 and 5,413,924 describe methods for preparing reagents for use in such methods. US Patent 6,403,341 describes a sequestering approach that involves chemical alteration of one of the PCR reagent components. In the most preferred embodiment, both RNA- and DNA-dependent polymerase activities reside in a single enzyme. Other components that are required for efficient amplification include nucleoside triphosphates, divalent salts and buffer components. In some instances, non-specific nucleic acid and enzyme stabilizers may be beneficial.

In the preferred RT-PCR, the amounts of certain reverse transcriptase and the PCR components are atypical in order to take advantage of the faster ramp times of some thermal cyclers. Specifically, the primer concentrations are very high.

Typical gene-specific primer concentrations for reverse transcriptase reactions are less than about 20 nM. To achieve a rapid reverse transcriptase reaction on the order of one to two minutes, the reverse transcriptase primer concentration is raised to

greater than 20 nM, preferably at least about 50 nM, and typically about 100 nM. Standard PCR primer concentrations range from 100 nM to 300 nM. Higher concentrations may be used in standard PCR to compensate for Tm variations. However, for the purposes herein, the referenced primer concentrations are for circumstances where no Tm compensation is needed. Proportionately higher concentrations of primers may be empirically determined and used if Tm compensation is necessary or desired. To achieve rapid PCR, the PCR primer concentrations typically are greater than 250 nM, preferably greater than about 300 nM and typically about 500 nM.

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Commercially used diagnostics also preferably employ one or more internal positive control that confirms the operation of a particular amplification reaction in case of a negative result. Potential causes of false negative results that must be controlled in an RT-PCR include: inadequate RNA quantity, degradation of RNA, inhibition of RT and/or PCR and experimenter error.

In the case of measuring protein levels to determine gene expression, any method known in the art is suitable provided it results in adequate specificity and sensitivity. For example, protein levels can be measured by binding to an antibody or antibody fragment specific for the protein and measuring the amount of antibody-bound protein. Antibodies can be labeled by radioactive, fluorescent or other detectable reagents to facilitate detection. Methods of detection include, without limitation, enzyme-linked immunosorbent assay (ELISA) and immunoblot techniques.

The invention provides specificity and sensitivity sufficient to identify a malignant melanocyte in a tissue sample. The methods determine expression of particularly Marker genes by measuring mRNA encoded by the Markers. The preferred Markers of the invention display at least a two-fold over-expression in tissue having malignant melanocytes relative to benign melanocyte or normal tissue. The results presented herein show that a primary Marker is insufficient to provide clinically relevant information but, when combined with one or more secondary Markers, the information obtained compares to the "gold standard" of H&E and IHC upon which clinicians currently rely. Tertiary Markers and control genes can augment the primary and secondary Markers to further increase specificity and/or sensitivity.

As described in the following Examples, the Markers were identified by the protocol depicted in Figure 1. Thus, the invention provides a method for identifying melanoma-specific Markers by following the protocol in Figure 1 and the Examples provided herein.

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The primary Marker can be PLAB and is defined herein as the gene encoding any variant, allele etc. including SEQ ID NO: 1. PLAB is also described by Paralkar et al. (1998) and represented by Accession No. AF003934. PLAB is linked to the pathogenesis of prostate cancer (Liu et al (2003); Karan et al. (2003); and Nakamura et al. (2003); US Patent Nos. 5,994,102; 6,107,476; 6,465,181; 6,500,638; 6,521,227; US Patent Publication Nos. 2002/0048784; 2003/0013097; and 2003/0059431) and colorectal cancer (Brown et al. (2003); Buckhaults et al. (2001); and US Patent Publication No. 2002/0160382).

The secondary Marker is LlCAM and is defined herein as the gene encoding any variant, allele etc. including SEQ ID NO: 2. LlCAM is also described by Haspel et al (2003); and US Patent Nos. 5,872,225; and 5,969,124 and is represented by Accession No. NM_000425.

The invention further provides tertiary markers that fall into several functional categories. Thus, additional Markers can be used that are found in these functional categories. As described in more detail in the Examples, melanoma-specific up-regulated genes fall into the functional categories of neural tissue development and cell cycle control and melanoma-specific down-regulated genes fall into the functional categories of tissue development and cell differentiation.

The tertiary Markers include SEQ ID NOs: 3, 29-978 and 999. A number of tertiary markers are described in Table 5 and all are summarized in Table 15.

NTRK3 is described by Strausberg et al. (2002); Marchetti et al. (2003); Hisaoka et al. (2002); McGregor et al. (1999); Ryden et al. (1996); US Patent Nos. 5,348,856; 5,844,092; 5,910,574; and US Patent Publication Nos. 2002/0155480; and 2003/014283 and is represented by Accession No. BC013693 or S76476.1. NTRK3 is also defined as the gene encoding mRNA recognized by the primer/probe sets SEQ ID NOs: 16-18.

Tyrosinase is described by Mandelcorn-Monson et al. (2003); and US Patent No. 6,153,388 and is represented by Accession No. NM_000372. Tyrosinase is also defined as the gene encoding mRNA recognized by the primer/probe sets SEQ ID NOs: 19-21.

5 Table 5

Gene Reference Accession of PBGD Raich et al. (1986) NM 00019 CITED1 Fenner et al. (1998) NM 00414 PEX6 Raas-Rothschild et al. (2002) NM 0028 CAPG Van Impe et al. (2003) NM 00174 DUSP4 Smith et al. (1997) NM 00139 GDF1 Ducy et al. (2000) NM 00149 E2-EpF Liu et al. (1992) NM 01450 me20m Maresh et al. (1994) U01874 CDH3 Patel et al. (2003) NM 00179 SMARCD3 Ring et al. (1998) NM 00307 PKM2 Luftner et al. (2003) NM 00265 GPI Tsutsumi et al. (2003) NM 00017 Pig10 Polyak et al. (1997) AF010413 CPEB1 Welk et al. (2001) NM 03059 HOXHB9 Catala et al. (2002) AI738662 Truncated calcium binding protein EST Project Hillier et al. (1995) NM 01327 SAAS Kikuchi et al. (2002) NM 00701 HCN2 Edgar et al. (2003) NM 00119 <td< th=""><th>0 3 7 7 4 2 1 3 8 4</th></td<>	0 3 7 7 4 2 1 3 8 4
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HCN2 Stieber et al. (2003) NM_00119	1
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MBP Kamholz et al. (1996) M13577	4
1111011	
AD3LPAD5 Li et al. (1995) U34349	
PLOD3 Wang et al. (2002) NM_00108	4
MC1R Salazar-Onfray et al. (2002)	
MIF Shimizu et al. (1999) NM_00241	5
HOXB7 Care et al. (1996) NM_00450	2
AIM1 Ray et al. (1997) XM_16630	0
EpHB6 Hafner et al. (2003) NM_00444	5
AKT1 Majumder et al. (2004) NM_00516	3
AKT2 Gosmanov et al. (2004) NM_00162	
AKT3 Xu et al. (2003) NM_00546	5
APH-1A Xu et al. (2003) NM 01602	
APP Masters et al. (1985) NM 20141	2
BACE Pastorino et al. (2004) NM_13897	
BACE2 Pastorino et al. (2004) NM_01210	4
CAPN1 Altznauer et al. (2004) NM 00518	3
CAPN2 Alexa et al. (2004) NM 00174	4 3 4

CDK5	Qi et al. (2004)	NM 004935
CDK5R1	Kam et al. (2004)	NM 003885
CSNK1A1	Burzio et al. (2002)	NM 001892
CSNK1D	Li et al. (2004)	NM 139062
CSNK1E	Swiatek et al. (2004)	NM 152221
CSNK2A1	Hilgard et al. (2004)	NM 001895
CSNK2A2	Szebeni et al. (2003)	NM 001896
CSN2K2B	Lee et al. (2004)	NM 001320
GSK3B	Chen et al. (2003)	NM 182946
MAPK1	Nishihara et al. (2004)	NM 138957
MAPK14	Bendotti et al. (2004)	NM 139014
MAPK3	Nishihara et al. (2004)	NM 002746
MAPT	Yu et al. (2004)	NM 016841
NCSTN	Shirotani et al. (2004)	NM 015331
PEN2	Marlow et al. (2003)	NM 172341
PRKACA	Sakwe et al. (2004)	NM 207518
PRKACB	Dwivedi et al. (2004)	NM_002731
PRKACG	Zhang et al. (2004)	NM 002732
PRKAR1A	Gronholm et al. (2003)	NM 212472
PRKAR2A	MacDougall et al. (2003)	NM 004157
PRKAR2B	Dwivedi et al. (2004)	NM 002736
PRKCE	Schechtman et al. (2004)	NM 005400
PSEN1	Pitsi et al. (2004)	NM 000021
PSEN2	Zatti et al. (2004)	NM 012486
PSFL	Clark et al. (2003)	NM 031301
ABL1	Gustafson et al. (2004)	NM 007313
ACK1	Ahmed et al. (2004)	NM 005781
ACTN4	Menez et al. (2004)	NM 004924
ARF1	Kadaja et al. (2004)	NM 001658
ARPC1B	Kaneda et al. (2002)	
BCAR3	Clark et al. (2003)	NM 003567
BRAF	Sasaki et al. (2004)	NM_004333
CDC42	Chen et al. (2004)	NM 044472
CRK	Stoletov et al. (2004)	NM_016823
CRKL	Zhang et al. (2003)	NM 005207
DDEF1	Oda et al. (2003)	NM_018482
DOCK1	Grimsley et al. (2004)	NM_001380
FYN	Lee et al. (2004)	NM_153048
GIT1	Haendeler et al. (2003)	NM_014030
GRB2	Zhou et al. (2004)	NM 203506
GRF2	Arozarena et al. (2004)	NM_006909
HRAS	Nomura et al. (2004)	NM_005343
JUN	Schmuth et al. (2004)	NM_002228
KRAS2	Qi et al. (2004)	NM_033360
MAP2K1	Rhee et al. (2004)	NM_002755

MAP2K2	Chen et al. (2004)	NM 030662
MAP2K4	Woo et al. (2004)	NM 003010
MAP3K11	Zhang et al. (2004a)	NM 002419
MAPK8	Fujii et al. (2004)	NM 139049
MYLK	Oury et al. (2004)	NM 053032
NRAS ·	Reifenberger et al. (2004)	NM 002524
PAK1	Sells et al. (1997)	HSU24152
PAK2	Kirchhoff et al. (2004)	NM_002577
PAK3	Kitano et al. (2003)	NM_002578
PAK4	Barac et al. (2004)	NM_005884
PAK6	Ching et al. (2003)	NM_020168
PAK7	Jaffer et al. (2002)	NM 020341
PTK2	Golubovskaya et al. (2004)	NM 005607
PXN	Saito et al. (2004)	NM_002859
RAC1	Pontow et al. (2004)	NM_198829
RAF1	Akula et al. (2004)	NM_002880
RAP1A	Nomura et al. (2004)	NM_002884
RAP2B	Evellin et al. (2002)	NM_002886
SHC1	Yannoni et al. (2004)	NM_183001
SOS1	Buchs et al. (2004)	NM_005633
SRC	Encinas et al. (2004)	NM_198291
TLN1	Tremuth et al. (2004)	NM_006289
VASP	Tokuo et al. (2004)	NM_003370
VCL	Izard et al. (2004)	NM_003373
WASPIP	Luthi et al. (2003)	NM_003387
ZYX	Li et al. (2004)	NM_003461

Tertiary Markers are able to replace and/or supplement primary or secondary Markers provided that the resulting assays have adequate sensitivity and specificity.

The specificity of any given amplification-based molecular diagnostic relies heavily, but not exclusively, on the identity of the primer sets. The primer sets are pairs of forward and reverse oligonucleotide primers that anneal to a target DNA sequence to permit amplification of the target sequence, thereby producing a target sequence-specific amplicon. The primers must be capable of amplifying Markers of the disease state of interest. In the case of the instant invention, these Markers are directed to melanoma.

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The reaction must also contain some means of detection of a specific signal. This is preferably accomplished through the use of a reagent that detects a region of DNA sequence derived from polymerization of the target sequence of interest. Preferred reagents for detection give a measurable signal differential when bound to a specific

nucleic acid sequence of interest. Often, these methods involve nucleic acid probes that give increased fluorescence when bound to the sequence of interest. Typically, the progress of the reactions of the inventive methods are monitored by analyzing the relative rates of amplicon production for each PCR primer set.

- 5 The invention further includes primer/probe sets and their use in the claimed methods. The sequences are:
 - SEQ ID NO:4 (PLAB forward primer) ggcagaatcttcgtccgca
 - SEQ ID NO: 5(PLAB reverse primer) ggacagtggtccccgttg
 - SEQ ID NO: 6 (PLAB probe) cccagctggagttgcacttgcggcc
- 10 SEQ ID NO:7 (PLAB upper primer) gaacaccgacctcgtccc
 - SEQ ID NO: 8 (PLAB lower primer) ggcggcccgagagata
 - SEQ ID NO:9 (PLAB probe) cgccagaagtgcggctgggattt
 - SEQ ID NO: 10 (LlCAM forward) gctgggactgggaacagaact
 - SEQ ID NO:11 (LICAM Reverse) ggagcagagatggcaaagaaa
- 15 SEQ ID NO:12 (LlCAM probe) ttcccaccatctgctgt
 - SEQ ID NO: 13 (L1CAM upper) ccacagatgacatcagcctcaa
 - SEQ ID NO:14 (LICAM lower) ggtcacacccagctcttcctt
 - SEQ ID NO: 15 (LlCAM probe) tggcaagcccgaagtgcagttcctt
 - SEQ ID NO: 16 (NTRK3 primer) gccccggcacccttta
- 20 SEQ ID NO:17 (NTRK3 primer) aaccetgccagtggtggat
 - SEQ ID NO: 18 (NTRK3 probe) cagatgggtgttttc
 - SEQ ID NO: 19 (Tyr upper) actcagcccagcatcattcttc
 - SEQ ID NO: 20 (Tyr lower) atggctgttgtactcctccaatc
 - SEQ ID NO:21 (Tyr probe) cttctcctcttggcagattgtctgtagctt
- 25 SEQ ID NO:22 (PBGD upper) ccacacacagcctactttccaa

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- SEQ ID NO:23 (PBGD lower) tacccacgcgaatcactctca
- SEQ ID NO:24 (PBGD probe) aacggcaatgcggctgcaacggcggaatt

Monitoring amplicon production may be achieved by a number of detection reagents and methods, including without limitation, fluorescent primers, and fluorogenic probes and fluorescent dyes that bind double-stranded DNA. Molecular

beacons, Scorpions, and other detection schemes may also be used. A common

method of monitoring a PCR employs a fluorescent hydrolysis probe assay. This method exploits the 5' nuclease activity of certain thermostable DNA polymerases (such as Taq or TfI DNA polymerases) to cleave an oligomeric probe during the PCR process.

The invention further provides amplicons obtained by PCR methods utilized in the inventive methods. These amplicons include the following:

SEQ ID NO:25 (PLAB Amplicon)

10 SEQ ID NO:26 (Ll CAM Amplicon)

ccacagatgacatcagcctcaagtgtgaggccagtggcaagcccgaagtgcagttccgctggacgagggatggtgtcca cttcaaacccaaggaagagctgggtgtgacc

SEQ ID NO:27 (tyrosinase Amplicon)

actcagcccagcatcattcttctcctcttggcagattgtctgtagccgattggaggagtacaacagccat

15 SEQ ID NO:28 (PBGD Amplicon)

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 ${\tt ccacacacagcctactttccaagcggagccatgtctggtaacggcaatgcggctgcaacggcggaagaaaacagcccaaagatgagagtgattcgcgtgggta}$

The oligomer is selected to anneal to the amplified target sequence under elongation conditions. The probe typically has a fluorescent reporter on its 5' end and a fluorescent quencher of the reporter at the 3' end. So long as the oligomer is intact, the fluorescent signal from the reporter is quenched. However, when the oligomer is digested during the elongation process, the fluorescent reporter is no longer in proximity to the quencher. The relative accumulation of free fluorescent reporter for a given amplicon may be compared to the accumulation of the same amplicons for a control sample and/or to that of a control gene, such as, without limitation, β-Actin or PBGD to determine the relative abundance of a given cDNA product of a given RNA in a RNA population. Products and reagents for the fluorescent hydrolysis probe assay are readily available commercially, for instance from Applied Biosystems.

Suitable detection reagents are commonly referred to as "Scorpions" and are described in US Patents 6,326,145 and 5,525,494. These reagents include one or more molecules comprising a tailed primer and an integrated signaling system. The

primer has a template binding region and a tail comprising a linker and a target binding region. The target binding region in the tail hybridizes to complementary sequence in an extension product of the primer. This target specific hybridization event is coupled to a signaling system wherein hybridization leads to a detectable change. In PCR the target binding region and the tail region are advantageously arranged such that the tail region remains single stranded, i.e. uncopied. Thus the tail region is non-amplifiable in the PCR amplification products. The linker comprises a blocking moiety that prevents polymerase mediated chain extension on the primer template.

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The most preferred detection reagents are TaqMan® probes (Roche Diagnostics, Branchburg, NJ) and they are described in US Patents 5,210,015; 5,487,972; and 5,804,375. Essentially, these probes involve nucleic acid detection by virtue of the separation of a fluor-quencher combination on a probe through the 5'-3' exonuclease activity of the polymerase used in the PCR. Any suitable fluorophore can be used for any of the Markers or controls. Such fluorophores include, without limitation, Texas Red, CaI Red, Fam, Cy3 and Cy5. In one embodiment, the following fluorophores correspond to the noted Markers: PLAB: Fam; LlCAM: Texas Red or CaI Red, tyrosinase: Cl; PBGD: Cy5.

Equipment and software also are readily available for controlling and monitoring amplicon accumulation in PCR and QRT-PCR including the Smart Cycler thermocylcer commercially available from Cepheid of Sunnyvale, California, and the ABI Prism 7700 Sequence Detection System, commercially available from Applied Biosystems.

In the case of gene expression assays, it is preferable to use a gene constitutively expressed in the tissue of interest. PBGD is commonly used as an internal control due to several factors: it contains no known pseudogenes in humans, it is constitutively expressed in human tissues and it is expressed at a relatively low level and therefore is less likely to cause inhibition of the amplification of target sequences of interest. Use of PBGD as a control minimizes or eliminates reporting erroneous results arising from all potential sources of false negative results.

In the commercialization of the described methods for QRT-PCR certain kits for detection of specific nucleic acids are particularly useful. In one embodiment, the kit includes reagents for amplifying and detecting Markers. Optionally, the kit includes sample preparation reagents and or articles (e.g., tubes) to extract nucleic acids from lymph node tissue. The kits may also include articles to minimize the risk of sample contamination (e.g., disposable scalpel and surface for lymph node dissection and preparation).

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In a preferred kit, reagents necessary for the one-tube QRT-PCR process described above are included such as reverse transcriptase, a reverse transcriptase primer, a corresponding PCR primer set (preferably for Markers and controls), a thermostable DNA polymerase, such as Taq polymerase, and a suitable detection reagent(s), such as, without limitation, a scorpion probe, a probe for a fluorescent hydrolysis probe assay, a molecular beacon probe, a single dye primer or a fluorescent dye specific to double-stranded DNA, such as ethidium bromide. The primers are preferably in quantities that yield the high concentrations described above. Thermostable DNA polymerases are commonly and commercially available from a variety of manufacturers. Additional materials in the kit may include: suitable reaction tubes or vials, a barrier composition, typically a wax bead, optionally including magnesium; reaction mixtures (typically 10X) for the reverse transcriptase and the PCR stages, including necessary buffers and reagents such as dNTPs; nuclease-or RNase-free water; RNase inhibitor; control nucleic acid(s) and/or any additional buffers, compounds, co-factors, ionic constituents, proteins and enzymes, polymers, and the like that may be used in reverse transcriptase and/or PCR stages of QRT-PCR. Optionally, the kits include nucleic acid extraction reagents and materials. Instructions are also preferably included in the kits.

The following examples are provided to illustrate but not limit the claimed invention. AU references cited herein are hereby incorporated herein by reference.

Example 1 Tissue Preparation

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Fresh frozen malignant melanoma, benign skin nevi, normal skin, melanoma lymph node metastasis and melanoma-free lymph node samples were obtained from Genomics Collaborative, Inc. (Cambridge, MA), Asterand (Detroit, MI), Clinomics (Pittsfield, MA) and Proteogenex (Los Angeles, CA), Ardais (Lexington, MA) and Impath (Westborough, MA). All tissue vendors declared that tissue specimens used in the study were collected according to an Institutional Review Board approved protocol of corresponding hospitals and principles of bioethics. Patients' demographic and pathology information was also collected. The histopathological features of each sample were reviewed to confirm diagnosis, and to estimate sample preservation and tumor content.

Melanoma and benign nevi primary tissues chosen for microarray analysis had melanocyte content greater than 50% with no mixed histology. Melanoma positive lymph nodes were collected from malignant melanoma patients; diagnosis of melanoma was confirmed by H&E in combination with IHC (SIOO and HMB45). Melanoma free lymph nodes derived from patients that did not have melanoma in their clinical history and absence of melanoma was confirmed by H&E and IHC using antibodies for SIOO and HMB45.

RNA from a total of 70 primary tissue samples was used for gene expression profiling and melanoma specific gene identification. Samples included 45 primary malignant melanoma, 18 benign skin nevi, and 7 normal skin tissues. The majority of primary melanomas included in the study represent early stage of disease and have thickness less than 4 mm, which is consistent with the standard melanoma patient population. Aitken et al. (2004). Patient demographic, clinical and pathology characteristics are presented in Table 6 and summarized in Table 7.

In addition, 77 malignant melanoma LN metastasis and 18 melanoma-free LN tissue samples were used for one-step quantitative PCR assay. Melanoma positive lymph nodes included axillary, cervical and inguinal lymph nodes with metastasis derived from epithelioid and spindle cell primary melanomas. Out of 18 melanoma free LN, 10 were collected from other cancer patients but no cancer cells were found in these nodes by pathologists and 8 LN were from non-malignant lesions.

Table 6

Sample ID	Age	Gen der	Race	Diagnosis	Location	T Stage	N&M Stage	Clark
430MM	n/a	F	Cau	normal skin	trunk			
431MM	n/a	F	Cau	normal skin	trunk			
432MM	n/a	F	Cau	normal skin	trunk			
433MM	n/a	F	Cau	normal skin	trunk			
435MM	n/a	F	Cau	normal skin	trunk			
437MM	n/a	F	Cau	normal skin	trunk		T	
485MM	37	М	Cau	normal skin	skin, NOS			
487MM	35	F	Cau	atypical nevus, mild	face			
489MM	56	F	Cau	compound nevus	face			
490MM	16	F	Cau	compound nevus	scalp & neck		Ī	
491MM	15	M	Cau	compound nevus	trunk			
493MM	35	F	Cau	compound nevus	trunk			
495MM	18	F	Cau	benign nevus, NOS	trunk			
496MM	21	F	Cau	intradermal nevus	lower limb & hip			
497MM	12	M	Cau	intradermal nevus	lower limb & hip			
498MM	44	F	Cau	benign nevus, NOS	trunk			
499MM	48	F	Cau	benign nevus, NOS	face			
500MM	65	М	Cau	intradermal nevus	trunk			
501MM	30	М	Cau	compound nevus	lower limb & hip			
502MM	20	F	Cau	compound nevus	trunk			
503MM	35	М	Cau	intradermal nevus	lower limb & hip			
504MM	23	М	Cau	compound nevus	trunk			
507MM	53	M	Cau	atypical nevus, moderate	trunk			
508MM	28	М	Cau	compound nevus	trunk			
509MM	43	М	Cau	intradermal nevus	trunk			
392MM	58	F	Cau	epithelioid melanoma	trunk	T3	NOMO	4
397MM	51	F	Cau	epithelioid melanoma	lower limb & hip	T2	NOMO	3
405MM	46	М	Cau	epithelioid melanoma	upper limb & shoulder	T2	NOMO	3
407MM	64	F	Cau	epithelioid melanoma	trunk	T1	NOMO	2
409MM	54	F	Cau	epithelioid melanoma	scalp & neck	T2	NOMO	3
440MM	61	М	Cau	malignant melanoma, NOS	lower limb & hip	T1	NOMO	2
441MM	78	М	Cau	spindle cell melanoma	face	T4	NOMO	5
442MM	52	М	Cau	malignant melanoma, NOS	upper limb & shoulder	T2	NOMO	3
443MM	51	F	Cau	spindle cell melanoma	trunk	T2	NOMO	3
444MM	49	F	Cau	spindle cell melanoma	lower limb & hip	T3	NOMO	4
445MM	76	F	Cau	malignant melanoma, NOS	upper limb & shoulder	Т3	NOMO	4
446MM	86	М	Cau	malignant melanoma, NOS	scalp & neck	T1	NOMO	2
447MM	48	М	Cau	epithelioid melanoma	skin, NOS	Т3	NOMO	4
448MM	72	F	Cau	epithelioid melanoma	upper limb & shoulder	T2	NOMO	3
449MM	62	М	Asian	epithelioid melanoma	lower limb & hip	Т3	N1M0	n/a
450MM	90	F	Cau	epithelioid melanoma	upper limb & shoulder	T4	N1M1	n/a
452MM	43	М	Cau	epithelioid melanoma	skin, NOS	Т3	NOMO	n/a
453MM	48	F	Cau	epithelioid melanoma	trunk	T3	NOMO	n/a
454MM	69	М	Cau	epithelioid melanoma	upper limb & shoulder	T3	Nomo	n/a
455MM	55	М	Cau	malignant melanoma, NOS	skin, NOS	T2	NOMO	n/a
456MM	63	М	Cau	malignant melanoma, NOS	lower limb & hip	T2	NOMO	3

457MM	69	M	Cau	spindle cell melanoma	trunk	T1	NOMO	2
459MM	86	F	Cau	malignant melanoma, NOS	lower limb & hip	T2	NOMO	3
460MM	64	М	Саи	malignant melanoma, NOS	upper limb & shoulder	ТЗ	NOMO	4
461MM	66	M	Cau	epithelioid melanoma	trunk	T1_	N0M0	2
463MM	58	M	Cau	malignant melanoma, NOS	trunk	T1	N0M0	2
464MM	53	M	Cau	epithelioid melanoma	face	T2	NOMO	3
465MM	77	F	Cau	epithelioid melanoma	upper limb & shoulder	ТЗ	NOMO	4
466MM	79	F	Cau	malignant melanoma, NOS	upper limb & shoulder	T1	N0M0	2
468MM	86	F	Cau	spindle cell melanoma	upper limb & shoulder	T2	NOMO	3
469MM	43	F	Cau	malignant melanoma, NOS	scalp & neck	T1	NOMO	2
470MM	81	М	Cau	malignant melanoma, NOS	upper limb & shoulder	T2	NOMO	3
472MM	38	F	Cau	spindle cell melanoma	upper limb & shoulder	T1	NOMO	2
473MM	69	F	Cau	malignant melanoma, NOS	upper limb & shoulder	T1	NOMO	3
475MM	77	F	Cau	malignant melanoma, NOS	face	Т3	NOMO	4
476MM	87	F	Саи	spindle cell melanoma	upper limb & shoulder	Т3	NOMO	4
477MM	82	М	Cau	malignant melanoma, NOS	scalp & neck	T2	NOMO	3
478MM	78 .	F	Cau	epithelioid melanoma	face .	T3	NOMO	4 .
480MM	59	М	Cau	malignant melanoma, NOS	upper limb & shoulder	T2	NOMO	3
481MM	85	М	Cau	malignant melanoma, NOS	upper limb & shoulder	ТЗ	NOMO	4
482MM	66	М	Cau	epithelioid melanoma	face	T3	NOMO	4
483MM	85	F	Cau	epithelioid melanoma	trunk	T4	NOMO	5
484MM	70	F	Cau	malignant melanoma, NOS	upper limb & shoulder	T1	NOMO	3
511MM	69	M	Cau	epithelioid melanoma	skin, NOS	T3	N1M0	4
512MM	45	M	Cau	epithelioid melanoma	trunk	T4	NOMO	3

Table 7

Characteristics	Melanoma (%)	Nevi (%)	Normal skin (%)
Mean Age	65.51 ± 14.55	33.17 ± 15.60	n/a
Gender			
Female	22 (48.9)	9 (50)	6
Male	23 (51.1)	0 (50)	1
Anatomical location			
Face	5 (11.1)	3 (17)	
Scalp and neck	4 (8.()	1 (6)	
Trunk	9 (20)	10 (55)	6 (86)
Upper limb and shoulder	17 (37.8)		
Lower limb and hip	6 (13.3)	4 (22)	
Skin, NOS	4 (8.9)		1 (14)
Histological diagnosis			
Epitheloid cell	20 (44.4)		
Spindle cell	7 (15.6)		

Malignant melanoma NOS	18 (40)		
Compound nevus	<u> </u>	8 (44)	
Intradermal nevus		5 (28)	
Atypical nevus		2 (11)	
Benign nevus, NOS		3 (17)	
Normal skin			7 (100)
T stage (thickness)			
T1	11 (24.4)		
T2	14 (31.1)		
T3	16 (35.6)		
T4	4 (8.9)		
N stage			
N0	42 (93.3)		
N1	3 (6.7)		
M stage			
M0	44 (97.8)		
M1	1 (2.2)		
Clark level			
2	9 (20)		
3	16 (35/6)		
4	12 (26.7)		
5	2 (4.4)		
n/a	6 (13.3)		

Example 2 RNA Isolation from malignant melanoma and benign skin nevi samples

Qiagen RNeasyTM Mini Kit (QIAGEN Inc., Valencia, CA) was used, with a 5 modified protocol to minimize the residual melanin in the RNA sample. For melanocyte containing tissues, four replicate tissue samples derived from individual patient each weighed approximately 5 mg and were used and processed separately. Tissue samples were homogenized in 1.0 ml RLT buffer (QIAGEN) containing 10 µl β-mercaptoethanol (Sigma Chemical Co., St. Louis, MO) by a mechanical 10 homogenizer (UltraTurrex T8, IKA-Werke, Staufen, Germany). After · homogenization, samples were loaded onto QIAGEN RNeasy™ columns and followed by centrifugation. After discarding the flow-through, 700 ml of RWl buffer was added; the column was kept for 5 min at room temperature and then centrifuged. This step was repeated 3 times. Then the standard QIAGEN RNeasyTM Mini Kit 15 protocol was followed. To remove RNA from the silica gel membrane, a two-step elution was performed. The total RNA derived from the same individual patient tissue was pooled and used for further analysis.

Standard Trizol protocol was used for RNA isolation from tissues that do not contain a significant proportion of melanocytes. Tissue was homogenized in Trizol reagent (Invitrogen, Carlsbad, CA). After centrifugation the top liquid phase was collected and total RNA was precipitated with isopropyl alcohol at -20°C. RNA pellets were washed with 75% ethanol, resolved in water and stored at -80°C until use. RNA quality was examined with an Agilent 2100 Bioanalyzer RNA 6000 Nano Assay (Agilent Technologies, Palo Alto, CA).

Labeled cRNA was prepared and hybridized with the high-density oligonucleotide array HuI 33A Gene Chip (Affymetrix, Santa Clara, CA) containing a total of 22,000 probe sets according to the standard manufacturer protocol. Arrays were scanned using Affymetrix protocols and scanners. For subsequent analysis, each probe set was considered as a separate gene. Expression values for each gene were calculated by using Affymetrix Gene Chip analysis software MAS 5.0. All chips met three quality control standards: "present" call was greater than 35%, scale factor was smaller than 12 when scaled to a target intensity of 600, and background level was less than 150. Lower than usual percent of "present" calls cut-off was chosen because it is difficult to isolate RNA from skin cells (Hipfel et al. (1998)) resulting in lower overall gene expression levels.

Example 3 Data Analysis

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Gene expression data were filtered to include only genes called "present" in 2 or more samples. This filter was used to remove genes that did not change expression in the samples. Of the 22,000 genes presented on the array, 15,795 passed this filter and were used for hierarchical clustering. Prior to clustering, each gene expression signal was divided by the median expression in al samples in the data set. This standardization step minimized the effect of the magnitude of gene expression and group together genes with similar expression patterns in the clustering analysis. Average linkage hierarchical clustering using Pearson correlation was performed on both the genes and the samples using GeneSpring 6.1.

In order to identify differentially expressed genes, we compared the melanoma samples to the benign nevi and the normal skin samples separately. The first analysis consisted of the 45 melanoma and 7 normal skin samples; the second analysis consisted of 45 melanoma and 18 nevi samples. These two datasets were analyzed separately in following procedures as shown in Fig. 1. Significance analysis of microarray (SAM; Tusher et al. (2001)) and Student T-test were used in gene selection. Parameters for SAM were set as Δ=2.5 and fold change = 2.0 with 1,000 permutations. FDR was 1%. There were no missing data and the default random number was used. Next percentile analysis was conducted. For up-regulated genes the 30%ile in melanoma samples was compared to the maximum of the normal samples, or that of nevi samples. Student T-test with Bonferroni correction was also performed with cut-off p<0.05 in order to ensure that the selected genes had significant differential expression between the two groups of the samples. As a final step, we identified common genes between the melanoma/benign and melanoma/normal gene lists resulting in the single list of genes upregulated in melanoma shown in Figure 1 where the 439 common genes correspond to SEQ ID NOs: 29-467 as described in Table 15 with the results shown in Table 8.

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Table 8					
	Median Expression in	Fold Change	Fold Change		
PSID	Melanoma	(Cancer vs Benign)	(Cancer vs skin)		
200078_s_at	3954	2	3		
200601_at	9254	2	7		
200612_s_at	2396	2	5		
200644_at	7240	3	6		
200660_at	14659	3	4		
200707_at	3153	2	51		
200736_s_at	7305	3	3		
200737_at	2423	2_	2		
200783 s_at	1028	2_	_ 2		
200825_s_at	3746	3	3		
200827_at	1593	2	2		
200837_at	5817	2	5		
200838_at	19225	8	17		
200839_s_at	28353	5	7		
200859_x_at	9665	3	7		
200910_at	7780	2	4		
200950_at	8419	3	7		
200954_at	3132	2	6		
200966_x_at	27388	2	3		
200967_at	6154	2_	4		
200968 s_at	5587	2	7		

200972_at	8943	3	3
201038_s_at	1480	2	2
201051_at	5439	2	_ 5
201105_at	33285	4	5
201106_at	7546	2	4
201188_s_at	1730	2	8
201189 s at	3870	3	3
201195 s at	6005	4	18
201202 at	1860	3	2
201251 at	23965	5	11
201252 at	901	2	3
201271_s_at	1648	2	5
201291 s at	601	3	42
201313 at	2414	5	6
201346 at	3359	2	2
201393_s_at	2166	2	5
201416 at	4845	4	4
201417 at	2905	2	4
201470 at	18525	4	4
201474 s at	2471	5	22
201474 s at	1398	2	10
201486_at	1105	2	3
201430 at	2441	2	4
201614 s at	651	2	4
201614 s at	4713	7	3
201661 s at	3336	6	5
201662 s at	2693	5	3
201670 s at	3047	2	12
2016/0 s at 201714 at	993	2	3
201765 s at	5475	4	5
201703_s_at 201792_at	3897	3	4
201792_at 201804 x at	5609	3	3
		5	3
201819_at	2575	10	20
201850_at 201880_at	11103 1833	2	
		4	3
201910_at	3854 2154		
201911 s_at		3	4
201931 at	2578	3	2
201954_at	25901	9	15
201976 s at	7697	2	6
202069_s_at	855	2	3
202070 s at	2490	4	5
202111_at	1380	3	9
202154_x_at	10260	3	4
202185_at	8493	4	6
202188_at	1269	3	9
202219_at	5630	3	7

202224_at	2650	3	4
202225_at	1534	2	2
202260 s_at	7877	6	6
202295 s at	15129	6	4
202329_at	3438	4	4
202367_at	898	2	2
202370 s at	4669	3	2
202478 at	7922	3	11
202503 s at	2424	3	5
202589 at	3494	2	6
202603 at	3772	3	2
202705_at	1058	4	4
202737 s at	2865	3	4
202779 s at	3400	9	55
202785 at	1134	2	6
202862 at	2540	6	4
202898 at	4736	4	63
202954 at	1357	2	2
202958 at	2401	4	3
202961_s_at	19648	4	5
202986 at	2052	3	28
203011 at	1346	2	2
203022 at ·	1068	2	2
203069 at	560	15	13
203071 at	958	19	3
203094 at	968	2	2
203145 at	513	2	2
203167 at	2523	3	5
203217 s at	6416	3	5
203234 at	725	3	3
203256 at	7799	5	31
203262 s_at	2482	2	2
203300_x_at	5278	3	16
203315_at	3273	2	2
203366_at	847	2	2
203396_at	3946	2	3
203452_at	579	4	12
203456_at	1132	2	3
203502_at	657	3	3
203518 at	2943	3	7
203554_x_at	5056	3	4
203557_s_at	839	2	2
203570 at	2553	4	15
203590 at	3093	3	7
203643_at	1380	3	8
203663 s at	9720	5	3
203668_at	1963	2 .	3

203693 s at	588	3	3
203695 s at	1548	6	3
203723 at	4087	5	9
203729 at	9154	2	14
203730 s at	517	3	3
203731 s at	691	3	2
203775 at	1251	4	3
203827_at	3380	11	8
203878 s at	2036	8	5
204014 at	7184	10	46
204015 s at	2207	5	—·
204013_s_at	T	7	15
	2154 664		7
204092 s at		3	3
204099 at	2496	6	6
204170 s at	1362	3	4
204197 s at	2889	3	4
204198_s_at	4024	3	3
204202_at	875	3	3
204228_at	1299	2	3
204244_s_at	538	4	4
204247 s at	689	3	27
204252_at	5294	5	7
204262 s at	1003	4	5
204423_at	689	4	3
204436_at	3113	3	3
204458_at	908	3	3
204467_s_at	1910	3	6
204584_at	9677	21	15
204585_s_at	805	13	17
204647_at	1693	3	3
204654 s at	3894	4	25
204709 s at	312	24	16
204778_x_at	537	2	44
204779_s_at	1641	4	4
204857_at	2736	3	297
204932_at	241	3	3
204973_at	1478	3	7
204995_at	496	4	_ 6
205051_s_at	3875	4	3
205142_x_at	937	2	3
205169 at	267	2	7
205373 at	579	10	8
205376 at	815	2	3
205405 at	2302	4	11
205447 s at	567	13	5
205458 at	3288	6	8
205566 at	2684	7	8
20000_at	2007	·	<u> </u>

205591_at	1190	5	3
205681_at	1316	8	12
205690_s_at	9179	8	9
205691_at	226	7	5
205717 x at	8127	3	3
205813_s_at	430	11	9
205937 at	301	7	5
205945_at	772	2	4
205996 s at	909	2	3
206128_at	364	3	3
206307 s at	534	8	4
206332_s_at	4671	2	2
206397_x_at	1436	6	43
206441_s_at	3681	6	31
206462_s_at	9953	53	24
206503_x_at	419	8	9
206617_s_at	898	4	19
206630_at	23194	3	46
206688_s_at	2989	3	2
206696_at	6446	7	191
206777_s_at	683	4	7
206864_s_at	421	5	5
206976 s at	3375	4	3
207038_at	1986	9	47
207060_at	497	4	5
207144 s at	593	17	24
207163_s_at	3217	3	10
207183_at	230	6	6
207592_s_at	350	5	14
207614_s_at	2139	2	6
207622 s at	882	2	16
207828_s_at	997	3	3
208002_s_at	3142	3	7
208089_s_at	1374	3	6
208308_s_at	12282	4	9
208540_x_at	5257	2	2
208644_at	2242	2	3
208657 s at	1547	2	4
208677_s_at	5414	3	14
208696_at	7351	4	3
208710_s_at	1112	3	44
208723_at	4402	3	6
208744 x at	1673	4	49
208837_at	3997	2	3
208916_at	1630	3	5
208928_at	1439	4	7
208956_x_at	7772	3	2

208974_x_at	6025	2	6
208975 s_at	1085	2	3
209015_s_at	1739	4	4
209036 s at	8944	3	3
209053 s_at	269	7	10
209072 at	6299	4	18
209079_x_at	12870	3	3
209081 s at	3160	3	2
209123 at	4686	4	3
209132 s at	4385	5	12
209172 s at	268	3	3
209197_at	820	3	3
209198 s at	491	3	3
209247 s at	1486	2	2
209254_at	1384	4	7
209255 at	4283	6	8
209256 s at	4949	8	6
209283 at	12529	5	3
209345 s at	1678	2	2
209407 s at	1461	2	6
209515 s at	5827	5	15
209773 s at	1243	3	5
209825 s at	765	2	3
209827 s at	4884	7	7
209828 s at	1146	4	5
209848 s at	32959	7	74
209875_s_at	3038	21	12
209932_s_at	7126	3	5
210052 s_at	1085	3	6
210073_at	337	4	6
210111_s_at	7841	5	3
210127_at	391	22	9
210854_x_at	2100	2	6
210926_at	574	2	3
210948_s_at	396	2	· 4
210951_x_at	2501	2	13
211013_x_at	498	8	13
211052 s_at	1399	3	2
211066_x_at	12431	2	2
211373 s at	2063	5	6
211564_s_at	1992	3	2
211752 s_at	2183	2	2
211759_x_at	5674	2	3
211833_s_at	502	2	28
212000_at	339	33	14
212070 at	13437	2	4
212081_x_at	1457	2	4

212119_at	4415	2	5
212178 s at	2976	3	14
212193_s at	3646	3	10
212247 at	1716	3	3
212285 s at	4252	2	4
212312 at	1234	4	2
212338 at	1598	3	4
212402 at	3019	3	4
212472 at	1987	5	5
212472 at	3747	5	4
212512 s at	1441	3	2
212520 s at	2188	2	4
212552 at	2611	2	2
		3	4
212715 s at	1085	-	
212739 s at	2736	2	3
212744_at	959	3	5
212745_s_at	376	2	15
212793_at	3123	4	5
212796 s at	2511	2	2
213002_at	1439	2	3
213007_at	968	3	4
213008_at	1086	6	10
213096_at	924	3	3
213131_at	. 1392	3	3
213169_at	4028	5	4
213215_at	1926	4	3
213217_at	5848	10	6
213241_at	9479	4	23
213274 s at	18263	12	26
213275_x_at	17604	7	3
213330 s at	1233	2	7
213333_at	1845	2	3
213338_at	932	6	4
213392_at	1022	2	2
213474_at	723	2	3
213496_at	2322	3	7
213573_at	1643	2	3
213587 s at	10416	18	9
213638_at	1827	26	102
213670_x at	1959	2	. 4
213720_s_at	2248	2	3
213746 s at	4187	3	14
213836 s at	2605	8	5
213895 at	1279	3	4
213960 at	11768	80	29
214023 x at	1602	7	9
214068_at	2148	9	10
214000_at	2140	<u> </u>	10

214104_at	814	2	3
214201_x_at	746	2	3
214581 x at	510	5	6
214614 at	542	10	9
214632 at	358	2	2
214656 x at	2977	2	2
214687 x at	26310	2	3
214708_at	249	2	3
214710 s_at	575	2	3
214714 at	2366	4	9
214717 at	471	4	4
214752 x at	6462	3	4
214778_at	311	3	27
214841 at	913	9	11
214893 x at	214	10	9
214896 at	3071	8	11
215025 at	2365	149	93
215115 x at	12421	34	15
215126_at	4940	8	19
215155_at	505	6	4
2153311 at	10093	86	30
215812_s_at	1176	3	13
215836 s at	9406	3	3
216194_s_at	5011	3	3
216973 s at	1732	6	4
217033_x_at	10961	21	19
217104_at	317	6	3
217226 s at	2191	3	3
217297 s at	838	3	21
217377 x at	12402	27	18
217419_x_at	2742	3	5
217624_at	349	21	20
217799_x_at	1724	2	11
217827 s at	4762	2	2
217867 x at	9024	3	9
217871 s at	19519	3	11
217891_at	1271	2	3
218009 s at	1557	4	3
218030 at	1316	2	3
218074 at	3594	2	4
218143 s at	4007	3	5
218151_x at	1384	2	2
218152 at	1440	2	3
218161_s_at	941	5	5
218175 at	3563	3	2
218330 s at	3853	7	4
		3	
218349_s_at	588		14

218359_at	796	3	5
218376 s at	1931	4	4
218447_at	2209	2	3
218542 at	409	3	6
218564_at	433	2	4
218608_at	627	3	4
218678 at	11356	14	20
218774_at	1061	2	5
218786 at	732	3	2
218824_at	1351	3	5
218839 at	1996	38	7
218856_at	3199	5	4
218888 s at	906	6	5
218931_at	911	3	2
218952_at	2661	7	7
218956_s_at	1840	4	_3
218980_at	1627	3	8
218996_at	1859	4	_6
219011_at	113	4	3
219039_at	1852	2	_5
219040_at	480	3	10·
219041_s_at	3435	4	2
219051_x_at	1127	3	8
219066_at	621	4	2
219143_s_at	3618	9	13
219148_at	426	2	3
219152_at	365	13	10
219219 at	859	2	3
219361_s_at	1033	3	7
219372_at	376	2	2
219408_at	421	3	36
219478_at	5485	80	18
219491_at	547	3	12
219522_at	822	4	3
219537_x_at	411	3	6
219555_s_at	402	12	21
219578_s_at	1419	17	26
219634 at	686	3	15
219637_at	355	3	3
219703_at	378	3	3
219742_at	409	3	9
219895_at	528	6	3
219933_at	1399	2	22
220116_at	748	5	8
220155_s_at	5010	5	6
220178_at	5915	9	15
220454_s_at	581	2	2

220864_s_at	8416	3	3
220948 s at	11794	2	3
220973_s_at	497	4	3
220974_x_at	2540	2	2
220980 s at	3598	3	2
221059_s_at	2438	5	3
221483 s at	9194	3	3
221484_at	3834	3	3
221538_s_at	3971	3	3
221558_s_at	2356	3	5
221577_x_at	4897	28	38
221641 s at	1199	2	4
221688_s_at	3740	2	4
221710_x_at	728	2	2
221732_at	931	3	2
221759_at	1261	4	21
221797_at	430	2	7
221799_at	1601	5	3
221815_at	3293	17	144
221882_s_at	1144	5	6
221902_at	2491	4	4
221909_at	243	22	17
221962 s at	1132	2	6
222116_s_at	4208	2	4
222153_at	445	3	8
222155_s_at	1264	3	10
222175_s_at	2415	3	6
222196_at	224	3	6
222199_s_at	2152	2	3
222206_s_at	383	4	12
222212 s at	3715	3	4
222231_s_at	2724	3	2
222234_s_at	754	4	12
222240_s_at	1331	3	3
222294_s_at	2193	3	5
32811_at	4194	2	3
40560_at	1629	3	5
44783_s_at	8503	14	6
46665_at	7835	4	3
55093_at	3032	4	4
63825_at	10096	14	74
87100_at	737	6	70

We selected a short list of genes with at least 10-fold over-expression in melanoma as compared to the benign specimens. The complete array dataset has been submitted to the NCBI/Genbank GEO database (series entry pending).

Hierarchical clustering revealed four distinct clusters (Fig. 2). Two clusters consisted of majority of the melanoma samples (43 out of 45); the third cluster included the majority of benign nevi samples (15 out of 18) and the fourth contained all 7 normal skin specimens. Melanoma samples themselves formed two clusters with 35 samples in one cluster and 10 samples in the other. Samples that formed the small cluster represented epithelioid melanoma only, visually contained less melanin and demonstrated higher expression of PRAME and MIA genes (p<0.05). The few stage III and IV tumors were all grouped in the small cluster. The large cluster showed higher expression of NTRK3 and nestin (NES) (p<0.05). All melanoma and benign nevi samples demonstrated equally high expression of known melanocyte markers such as tyrosinase and MART-I, confirming that there is comparable melanocyte content in these samples. Our data indicate that melanoma, benign nevi and normal skin samples have distinct gene expression profiles and can be separated on molecular basis. Selected genes that were highly expressed in melanoma and their associated functional categories are summarized in Table 9.

Table 9

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1 auto 9				
psid	Name	Median	Median	Fold change
		expression in	expression in	ļ
_		melanoma	benign/normal	
Neural system	n development and	l function		
215025_at	NTRK3	2365.1	19.9	118.8
204709_s_at	KNSL5	311.8	13.5	23.1
204585_s_at	L1CAM var 1	805.2	49	16.4
218678_at	NES	11355.5	703.6	16.1
202260 s at	STXBP1	7877.1	1312	6.0
204995_at	p35	496.3	89.2	5.6
208308_s_at	GP1	12281.7	2238.2	5.5
201340 s at	ENC1	390.8	74.9	5.2
209072_at	MBP	6299.1	1215.9	5.2
Cell moveme	nts			
214614_at	HOXB9	541.6	54.4	10.0
205447_s_at	MAP3K12	566.5	76.5	7.4
Tissue morphology				
206397_x_at	GDF1	1436.1	130.1	11.0

205458_at	MC1R	3287.6	458.1	7.2
Cancer cell in	ıvasion			<u> </u>
213274_s_at	CSTTB	18262.9	1261.4	14.5
208677_s_at	BSG	5413.8	1088	5.0
Cell cycle cor	itrol			
219578_s_at	CPEB1	1418.5	75.6	18.8
207144_s_at	CITED1	593.4	33.5	17.7
204252_at	CDK2	5293.7	869.3	6.1
211373_s at	PSEN2	2063.3	403.2	5.1
Cell death				
221577_x_at	PLAB	4896.9	173.9	28.2
205681_at	BCL2A1	1316.4	135.1	9.7
Unknown				
204545_at	PEX6	379.1	23.9	15.9
201850_at	CAPG	11103.2	725.6	15.3
204014_at	DUSP4	7183.6	601.3	11.9
202779_s_at	E2-EPF	3400	323.3	10.5
201954_at	ARPC1B	25900.7	2470	10.5
209848_s_at		32958.9	3778.4	8.7
213112_s_at	SQSTM1	260.4	33.9	7.7
218952_at	SAAS	2660.7	368.7	7.2
204099_at	SMARCD3	2496.2	428.2	5.8
206999_at	IL12RB2	354	61.3	5.8
201251_at	PKM2	23964.7	4228.2	5.7
202185_at	PLOD3	8493.2	1541.7	5.5

Example 4 Identification of genes differentially expressed in melanoma

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A total of 70 gene expression profiles were used for analysis. The he median percentages of "present calls" for melanoma, benign and normal sample groups were 43.8%, 46.9% and 41.7%. Sixty microarrays (86%) had scaling factors within 3-fold range of the minimum value. Ten chips with the scaling factors more than 3 were equally distributed between the sample categories, melanoma, benign and normal.

Unsupervised hierarchical clustering result revealed a distinct separation of the melanoma, benign nevi and normal skin samples (Fig. 2). We observed four clusters, including two clusters consisting of majority of the melanoma samples (43 out of 45), the third cluster contained all 7 normal skin, 3 benign nevi and 2 melanoma specimens and the fourth cluster, that included 14 of the 18 benign nevi samples. Source of the samples did not affect clustering. Specimens originated from different sources were clustered together according the sample type (melanoma, benign or

normal). To further test the stability of the clustering patterns, we used an alternative cut-off on gene filtering prior to the cluster analysis. Specifically, we retained genes that have at least 10% "present" calls in each of the melanoma, benign nevi and skin samples. With this cut-off, we obtained 15, 306 genes and repeated hierarchical clustering. The cluster pattern on the patient samples was the same as the one from the 15,795 from the 2 "present" calls, confirming clustering stability.

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The single nevi sample that clustered with the melanoma samples is an atypical nevi (moderate degree) sample with no melanoma *in-situ* present. All three nevi samples that clustered with normal skin are compound nevi samples and one of them has melanocyte content lower than the other nevi specimens. The melanoma samples themselves formed two clusters with 34 samples in the large and 9 samples in the smaller cluster. Samples that formed the small cluster represented epithelioid melanoma only and visually contained less melanin. The few stage III and IV tumors, used in our study, were all grouped in the small cluster. The large cluster was composed from epithelioid, spindle cell and melanoma of mixed histology specimens with more significant presence of melanin. The large cluster included Stage I and Stage II specimens only.

Distinct gene clusters were found in association to melanoma. This can be characterized by up-regulated (Fig. 2, A, B, C) and down-regulated (Fig. 2, E) genes in the melanoma samples. At the same time, melanoma and benign nevi samples demonstrated high expression of known melanocyte markers, such as MART-I (Fig. 3, D) confirming a comparable content of melanocyte in these samples and inability of melanocyte specific markers to differentiate them. Our data indicate that melanoma, benign nevi and normal skin samples have distinct gene expression profiles and can be separated on their molecular basis.

In order to identify genes upregulated in malignant melanoma, we applied SAM in combination with t-test with Bonferroni correction and percentile analysis (Fig. 1). Bonferroni-adjusted t-test and percentile analyses were used to address the multiple testing issue and the heterogeneity of the tumor samples, respectively. As the result of these analyses, 439 genes were selected and are summarized in Table 15 as SEQ ID NOs: 29-467. Out of 439 genes up-regulated in melanoma, we selected a short list of

WO 2006/002433 PCT/US2005/022846

33 genes that had more than 10-fold over-expression in the melanoma samples than that of the benign specimens. These include many genes with known association with malignant melanoma such as NTRK3 (Xu et al. (2003)), LlCAM (Fogel et al. (2003); and Thies et al. (2002)), me20m (Adema et al. (1994)), as well as novel genes. Genes with more than 10-fold overexpression in melanoma are presented in Table 10.

Table 10

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Psid	Description	Median Exp	Fold change	Fold change
		Melanoma	(Can v Benign)	(Can v skin)
215025_at	NTRK3	2365	149	93
215311_at	EUROIMAGE 21920	10093	86	30
213960_at	EUROIMAGE 51358	11768	80	29
219478_at	WFDC1	5485	80	18
218839_at	HEY1	1996	38	7
215115_x_at	TEL oncogene	12421	34	15
221577_x_at	PLAB	4897	28	38
217377 x at	ETV6-NTRK3 fusion	12402	27	18
213638 at	PHACTR1	1827	26	102
204709_s_at	KNSL5	312	24	16
221909_at	Hyp protein FLJ14627	243	22	17
204584_at	L1CAM	9677	21	15
209875_s_at	SPP1	3038	21	12
217624_at	PDAP1	349	21	20
203071 at	SEMA3B ·	958	19-	3 .
213587 s_at	C7ORF32	10416	18	9
221815_at	ABHD2	3293	17	144
219578_s_at	CPEB1	1419	17	26
207144_s_at	CITED1	593	17	24
203069_at	SV2A	560	15	13
218678_at	NES	11356	14	20
219152_at	PODXL2	365	13	10
205447_s_at	MAP3L12	567	13	5
213274_s_at	CTSB	18263	12	26
219555_s-at	BMO39	402	12	21
203827_at	WIPI49	3380	11	8
205813_s_at	MAT1A	430	11	9
201850_at	CAPG	11103	10	20
205373_at	CTNNA2	579	10	8
214614_at	HLXB9	542	10	9
213217	ADCY2	5848	10	6
204014_at	DUSP4	7184	10	46
214893	HCN2	214	10	9

We further selected three genes over-expressed in melanoma, including NTRK3, PLAB, LlCAM, for quantitative real-time RT-PCR validation of the microarray results (Fig. 3). PLAB is a novel gene, whose differential expression in melanoma was not reported before at our best knowledge. For LlCAM and NTRK3, differential

expression in melanoma was demonstrated at protein level only. Xu et al. (2003); Fogel et al. (2003); and Thies et al. (2002). Moreover, we identified PLAB and LICAM as the best combination, on complementary basis, to separate melanoma from benign/normal tissues in our study. GPIOO is known as a melanoma specific marker and was selected as positive control. For the RT-PCR assay we used a panel of 14 primary melanoma, 7 benign nevi and 5 normal skin samples, isolated from the same tissues as used for the microarray study. The expression value of each gene was normalized to the housekeeping control gene PBGD. The correlation coefficients between the RT-PCR and the microarray results for LICAM, NTRK3, PLAB and gpIOO are 0.79, 0.86, 0.87 and 0.88, respectively. This result indicates that the RT-PCR results are highly consistent with the microarray data.

Example 5

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Pathway Analysis of Differentially Expressed Genes

Functional analysis of genes differentially expressed in melanoma was performed using IngenuityTM Pathway Analysis Software Application (Ingenuity, Mountain View, CA). Functional categories or canonical pathways that have p-value of less than 0.05 were selected. Specificity of canonical pathways identification was tested using randomly selected genes.

In order to gain further insight into a potential mechanism that differentiates melanoma from benign and normal tissue, we used Ingenuity pathway analysis software to identify canonical pathways associated with melanoma. The results analysis revealed that many of the genes in amyloid processing were up-regulated in the melanoma samples. To verify specificity of our observation, we selected three random lists of genes from Affymetrix HuI 33A microarray and subjected them to Ingenuity pathway analysis. None of these lists produced a significant association to amyloid processing or any other canonical pathways. To confirm the activation of this canonical pathway in melanoma, gene expression data for all the genes in the pathway were retrieved. Fold-change and p-value of differential expression between melanoma and benign/normal tissues were calculated. Out of the 34 genes included in the amyloid processing pathway (Esler et al. (2001); and Giancotti et al. (1999)), 25 demonstrated up-regulation trend and for 19 of them (56%), differential expression was statistically significant (p-value< 0.05; Fig 4). As the additional control, we

randomly selected two metabolic pathways with a similar number of genes. Out of the 63 genes in alanine synthesis pathway, 8 of them (13%) showed significant upregulation with p-value less than 0.05. Out of the 47 genes in histidine synthesis pathway, only 2 genes (4%) were found using the same criteria. For the first time, our data strongly indicated that activation of the amyloid processing pathway is involved in malignant melanoma.

Example 6 RT-PCR Validation of Microarray Results

Ten microgram total RNA from each sample was treated with DNase I and reverse-transcribed with oligo (dT) primer using Superscript II reverse transcriptase according to the manufacturer's instructions (Invitrogen, Carlsbad, CA). A control gene PBGD was previously tested and reported as a housekeeping gene.

Vandesompele et al. (2003). Primers and MGB-probes for me20m (gplOO), LlCAM, NTRK3, and the control gene PBGD were designed using Primer Express software (Applied Biosystems, Foster City, CA). The PLAB (MICl) gene probe was FAM-TAMRA based since sequences were inadequate to design MGB based probes.

Primer/probe sequences were as follows:

Table 11

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Description	Sequence	SEQ ID NO:
me20m forward	TGTGTCTCTGGCTGATACCAACA	983
me20m reverse	TTCTTGACCAGGCATGATAAGCT	984
me20m probe	(6-FAM) CTGGCAGTGGTCAGC	985
L1CAM forward	GCTGGGACTGGGAACAGAACT	10
L1CAM reverse	GGAGCAGAGATGGCAAAGAAA	11
L1CAM probe	(6-FAM) TCCCCACCATCTGCTGT	12
NTRK3 forward	GCCCGGCACCCTTTA	16
NTRK3 reverse	AACCCTGCCAGTGGTGGAT	17
NTRK3 probe	(6-FAM) CAGATGGGTGTTTTC	18
PLAB forward	GGCAGAATCTTCGTCCGCA	4
PLAB reverse	GGACAGTGGTCCCCGTTG	5
PLAB probe	(6-FAM) CCCAGCTGGAGTTGCACTTGCGGCC(TAMRA)	6

PBGD forward	CTGCTTCGCTGCATCGCTGAAA	986
PBGD reverse	CAGACTCCTCCAGTCAGGTACA	987
PBGD probe	(6-FAM) CCTGAGGCACCTGGAAGGAGGCTGCAGTGT(TAMRA)	988

All primers and probes were tested for optimal amplification efficiency above 90%. The standard curve was composed of six 10-fold dilutions of target gene PCR product with copy numbers ranging from 10 to 10⁶. RT-PCR amplification was carried out in a 20μl reaction mix containing 50ng template cDNA, 2 x TaqMan[®] universal PCR master mix (12.5μl) (Applied Biosystems, Foster City, CA), 50OnM forward and reverse primers, and 25OnM probe. Reactions were run on an ABI PRISM 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA). The cycling conditions were: 2 min of AmpErase UNG activation at 50°C, 10 min of polymerase activation at 95°C and 50 cycles at 95°C for 15 sec and annealing temperature (60°C) for 60 sec. In each assay, a standard curve and a no-template control along with template cDNA were included in duplicate for both the gene of interest and the control gene. The relative quantity of each target gene was represented as ΔCt, which is equal to Ct of the target gene subtracted by Ct of the control gene.

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To confirm the melanoma specific genes identified by the microarray analysis, four genes (LlCAM, NTRK3, PLAB and gplOO) were selected for quantitative real-time RT-PCR validation (Fig. 4). The expression value of each gene was normalized to housekeeping control PBGD. The correlation coefficient between the RT-PCR and the microarray results for LlCAM, NTRK3, PLAB and gplOO are 0.79, 0.86, 0.87 and 0.88, respectively, indicating that the RT-PCR results are highly consistent with the microarray data.

Example 7 One-step qRTPCR Assays Using RNA-specific Primers and Cutoff Establishment

Evaluation of expression of selected genes was carried out with one-step RT-PCR with RNA from primary melanoma, benign nevi, normal skin, melanoma LN metastasis and melanoma-free lymph nodes. Beta-actin was used as a housekeeping gene to control for the input quantity and quality of RNA in the reactions. DNase

treatment was not used. Instead, primers or probes were designed to span an intron so they would not report on genomic DNA. Eight ng of total RNA was used for the RT-PCR. The Total RNA was reverse transcribed using 40X Multiscribe and RNase inhibitor mix contained in the TaqMan® One Step PCR Master Mix Reagents Kit (Applied Biosystems, Foster City, CA). The cDNA was then subjected to the 2x Master Mix without UNG and PCR amplification was carried out on the ABI 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA) in the 384-well block format using a 10 µl reaction size. The primer and probe concentrations were 4 µM and 2.5 µM, respectively. The reaction mixture was incubated at 48°C for 30 min for the reverse transcription, followed by a Amplitaq activation step of 95°C for 10 min and finally 40 cycles of 95°C for 15 sec denaturing and 60°C for 1 min anneal and extension. On each plate a standard curve is generated from 8 pg to 80 ng and when the R2 value was greater than 0.99 the Cycle Threshold (Ct) values were accepted.

Sequences used in the reactions were as follows, each written in the 5' to 3' direction.

Table 12

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Description	Sequence	SEQ ID NO:
L1CAM Forward	CCACAGATGACATCAGCCTCAA	13
L1CAM Reverse	GGTCACACCCAGCTCTTCCTT	14
L1CAM probe	TGGCAAGCCCGAAGTGCAGTTCC	15
Tyrosinase Forward	CTTTAGAAATACACTGGAAGGATTTGCTA	1000
Tyrosinase Reverse	CATTGTGCATGCTTTGA	1001
Tyrosinase probe	TCCACTTACTGGGATAGCGGATGCCTC	1002
MART1 Forward	ACTTCATCTATGGTTACCCCAAGAA	1003
MART1 Reverse	TCCCAGCGGCCTCTTCA	1004
MART1 Probe	CACGGCCACTCTTACACCACGGC	1005
HMB45 Forward	CTTAAGGCTGGTGAAGAGACAAGTC	1006
gp100 Reverse	CAGGATCTCGGCACTTTCAATAC	1007
gp100 Probe	TCGATATGGTTCCTTTTCCGTCACCCTG	1008
PLAB Forward	ATTCGAACACCGACCTCGTC	1009

PLAB Reverse	CGCAGGTGCAGGTGGC	1010
PLAB Probe	GATACTCACGCCAGAAGTGCGGCT	1011

For each sample Δ Ct=Ct (Target Gene) - Ct β -actin was calculated. Δ Ct has been widely used in clinical RT-PCR assays and was chosen as a straightforward method. Cronin et al. (2004). T-test was performed on Δ Ct between the melanoma and non-melanoma samples including both primary and LN samples. We then used Δ Ct to construct two scores for each patient. One score was derived from a combination of 2 melanoma specific genes, PLAB and LlCAM; and the other score was derived from a combination of 3 conventional melanoma markers, tyrosinase, gplOO and MARTl. The score was defined as the weighted sum of Δ Ct values of the tested genes with the corresponding t statistics as the weight. The two scores were normalized to have the same mean in order to compare them on the same scale.

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We examined a combination of two highly overexpressed in melanoma genes, PLAB and LICAM, in a variety of clinical tissue samples containing malignant melanocytes (primary melanoma and melanoma LN metastasis), benign melanocytes (benign skin nevi) and normal samples (normal skin and melanoma- free LN) by RT-PCR. The primary tissues were the same as those used for the microarray study while all the LN specimens were derived from independent patients. Conventional melanoma markers, such as tyrosinase, gplOO and MARTl, were also tested on the same samples as the controls because they are the most commonly used markers for the melanoma molecular assays in current clinical studies. Rimboldi et al. (2003); Abrahamsen et al. (2005); and Kammula et al. (2004). Calculated scores were presented on Fig. 4A for PLAB and LlCAM and on Fig. 4B for tyrosinase, gplOO and MARTI. The results demonstrated significant difference in expression of PLAB and LICAM between malignant melanoma samples (primary and LN metastasis) and benign nevi and normal LN. In contrast, three conventional markers showed similar expression levels in benign and melanoma samples. To further demonstrate the ability of gene markers to separate benign and malignant tissues, we tested two cutoffs; first was set up as the highest score in primary normal samples and the second as the highest score in benign nevi samples. For each cut-off we estimated sensitivity and of the assay in the LN samples. With the cut-off determined on the normal

samples, the new markers and the conventional markers gave sensitivity of 90% and 83%, respectively. Using the cut-off determined on the benign samples, the sensitivity for the new and conventional markers were 88% and 42%. The results indicated that the new markers potentially have better abilities to differentiate tissues containing benign and malignant melanocytes.

Example 8

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Multiplex Assay

Materials and methods

Each reaction was set up in a final volume of 25 µl containing the following:

10	forward primer	400 nM
	reverse primer	500 nM
	PLAB probe	150 nM
	Tyrosinase probe	300 nM
	LICAM probe	20O nM
15	PBGD probe	200 nM
	Tth	5 U
	Ab TP 6-25	iμg
	Glycerol	10%
	Tris-HCl	3J mM
20	NaCl	4 mM
	EDTA	0.004 mM
	Tween-20	0.22%
	NP-40	0.02%
	DTT	0.04 mM
25	Potassium Hydroxide	20.5 mM
	Bicine	50mM
	Potassium Acetate	115 mM
	Albumin, bovine	5 μg
	Trehalose	0.15 M
30	dNTP	0.2 mM ea
	MgCl ₂	0.5 mM
	MnSO ₄	3.5 mM
	Primers	300 nM ea
	Probes	200 nM ea

35 The primer and probe sequences are provided in Table 13.

Table 13

SEQ ID NO	Sequence 5'-3'	Function
43	gaacacegacetegtece	PLAB Upper Primer
44	ggcggcccgagagata	PLAB Lower Primer
45	Fam-cgccagaagtgcggctgggat-BHQ1-tt	PLAB Probe
55	acteageecageateattette	Tyr Upper Primer
56	atggctgttgtactcctccaatc	Tyr Lower Primer
57	Q570-cttctcctcttggcagattgtctgtagc BHQ2-tt	Tyr Probe
49	ccacagatgacatcagcctcaa	L1CAM Upper Primer
50	ggtcacacccagctcttcctt	L1CAM Lower Primer
51	CalRed-tggcaagcccgaagtgcagttcc-BHQ2-tt	L1CAM Probe
58	ccacacacagcctactttccaa	PBGD Upper Primer
59	tacccacgcgaatcactctca	PBGD Lower Primer
60	Q670-aacggcaatgcggctgcaacggcggaa-BHQ2-tt	PBGD Probe

The reactions are run with PLAB in Fam, Tyrosinase in Cy3, LlCAM in Texas Red, and PBGD in Cy5 channels. The cycling protocol used is described below and takes 30 min to complete.

5 95°C x 15 sec 65°C x 420 sec 40°cycles of:

95°C for 5 sec

62°C for 15 sec - fiuor read

The thresholds used are 30 in Fam, 20 in Cy3, 20 in Texas Red, and 20 in Cy5 channels. The thresholds employed in the Cy3 and Texas red channels can be lowered. The results obtained are summarized in Table 14.

Table 14
Best Marker Combinations

Markers	% Sensitivity (95% CI)	% Specificity (95% CI)
L1CAM + PLAB	82 (73-89)	96 (87-100)
Tyrosinase + ME20M (GP100)	63 (52-72)	100 (94-100)
L1CAM + PLAB + Tyrosinase	87 (79-93)	96 (87-100)

1'5 Ct cutoffs:

 LICAM
 27

 PLAB
 29

 Tyrosinase
 23

 ME20M (GPIOO)
 23.5

Note: these data are benchmarked against H&E pathology only. The amplification efficiency in each of the 4 reactions is high and the reaction is also linear over 5 logs (as judged by the R2 value which is >0.99 in all cases). Therefore,

these data demonstrate a working 4 plex, rapid assay. These data suggest that PLAB is the primary marker and complementation, achieved with LlCAM, further increases sensitivity. If required, addition of tyrosinase as a third marker further complements LlCAM and PLAB and increases sensitivity. Tyrosinase can be dropped from the assay, if needed, without affecting the performance of the remaining markers.

Discussion

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We performed gene expression profiling analysis of primary melanoma, benign nevi and normal skin tissue specimens in order to find melanoma specific gene markers for potential use in the LN molecular staging assay. Novel genes that are highly and differentially expressed in malignant melanoma samples were identified. Inclusion of benign nevi in the experimental design was key to our study. In contrast to normal skin, melanocyte content in benign nevi is close to that in melanoma. This was confirmed, in addition to histological assessment, by equally high expression level of conventional melanoma markers such as tyrosinase and MARTl in both melanoma and nevi tissue specimens. Similar cellular composition allowed us to monitor gene expression changes specifically associated with melanocyte malignant transformation, not just with melanocyte lineage differentiation. As the result, we identified novel genes specifically overexpressed in melanoma. One of the novel highly overexpressed in melanoma genes, prostate differentiation factor (PLAB, MICl), is a member of transforming growth factor-beta superfamily and also known to be associated with other malignancies. Bae et al. (2003); and Welsh et al. (2003). PLAB reduces cell adhesion (Yamauchi et al. (2003)), implicating its potential role in melanoma progression. Pathway analysis of the overexpressed genes in melanoma indicated that many of these genes belong to neural tissue functioning and development, suggesting that dedifferentiation of melanocytes and activation of the processes related to a pluripotent progenitor cell might be important for melanoma development and progression. Moreover, the analysis of canonical pathways showed that neural tissue associated amyloid processing is significantly modulated in melanoma. Amyloid processing (APP) pathway itself has not been associated with melanoma development and progression before. Many genes in the APP pathway, such as members of the β- and Y-secretase family (BACE2, PSEN2) also participate

in the Notch pathway and play a role of cleavage of integral membrane proteins in both Notch and APP. Esler et al. (2001). Notch suppresses differentiation and helps maintain neural crest stem cells in undifferentiated state (Gangemi et al. (2004)) and Notch's involvement in melanoma and, particularly, the role of Y-secretases is the focus of many studies. Hoek et al. (2004); Baldi et al. (2003); and Wilson et al. (2000).

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We have compared our results to the recent study of Haqq et al (2005). In their work, cDNA microarray containing 20,862 probes was used to profile benign nevi, primary melanoma and metastatic melanoma specimens. The sample set included metastatic and primary melanoma and benign nevi. Similar clustering results that separated the benign nevi and the primary malignant melanoma tissues were found in their study. Common genes were reported in both studies that can discriminate melanoma from benign nevi including kinesin-like 5 (KNSL5), prostate differentiation factor (PLAB), CITED1, osteopontin (SPPI), cathepsin B (CSTB), cadherin 3 (CDH3), presenilin 2 (PSEN2).

Our results of the one-step RT-PCR assay demonstrated that novel melanoma specific gene PLAB and LlCAM expressed not only in primary melanoma tissues but also in melanoma LN metastasis. Moreover, the ability to differentiate malignant melanoma from benign nevi made them better candidates than the conventional markers for the molecular test of melanoma diagnostics. With further validation in clinical studies, these genes could be developed as specific markers for a molecular staging assay to detect melanoma micrometastasis during sentinel lymph node (SLN) biopsy procedure. Another potential application of the genes is for diagnosis of melanocyte lesions with uncertain pathological features.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, the descriptions and examples should not be construed as limiting the scope of the invention.

Table 15 Sequence Descriptions, names and SEQ ID NOs:

	T	le Descriptions,	PLAB	
1		 		
3			L1CAM NTRK	
3	 		NIKK	DI AD formed primer
5				PLAB forward primer
	 	-		PLAB reverse primer
6		 		PLAB probe
7				PLAB upper primer
8				PLAB lower primer
9				PLAB probe
10				L1CAM forward primer
11				L1CAM reverse primer
12				L1CAM probe
13				L1CAM upper primer
14				L1CAM lower primer
15				L1CAM probe
16				NTRK primer
17			_	NTRK primer
18				NTRK probe
19				Tyrosinase upper primer
20			1	Tyrosinase lower primer
21				Tyrosinase probe
22				PBGD upper primer
23				PBGD lower primer
24				PBGD probe
25				PLAB amplicon
26				L1CAM amplicon
27				Tyrosinase amplicon
28	200078 s at	DC005076		PBGD amplicon
29	200078 s at	BC005876		ATPase, H+ transporting, lysosomal
30	200601_at	U48734 NM 001282	AP2B1	non-muscle alpha-actinin
31 32	200612_s_at	NM_023009	MACMARCKS	adaptor-related protein complex 2, β 1
32		14141_023009	IVIACIVIARCES	macrophage myristoylated alanine- rich C kinase substrate
33	200660_at	NM_005620	S100A11	S100 calcium-binding protein A11
34	200707_at	NM_002743	PRKCSH	protein_kinase C substrate 80K-H
35	200736_s_at	NM_000581	GPX1	glutathione_peroxidase_1
36	200737_at	NM_000291	PGK1	phosphoglycerate kinase 1
37	200783_s_at	NM_005563	LAP 18	leukemia-assoc phosphoprotein p18
38	200825_s_at	NM_006389	ORP150	oxygen_regulated_protein_(150kD)
39	200827at	NM_000302	PLOD	procollagen-lysine, 2-oxoglutarate 5- dioxygenase
40	200837 at	NM 005745	DXS1357E	accessory proteins BAP31BAP29
41	200838 at	NM 001908	CTSB	cathepsin B
42	200839 s at	NM 001908	CTSB	
43	200859_x_at	NM 001456	FLNA	filamin A, alpha
44	200910_at	NM 005998	CCT3	chaperonin containing TCP1, sub 3(γ)
45	200950at	NM_006409	ARPC1A	actin related protein 23 complex, sub
46	200950 at	NM 006409	ARPC1A	"
47	200956_x at	NM 000034	ALDOA	aldolase A, fructose-bisphosphate
48	200967 at	NM 000942	PPiB	peptidylprolyl isomerase B
49	200968_s_at	NM 000942	PPIB	popularity identifiade b
50	200900 s at	BC000704	1110	tetraspan 3
- 50	1 20001 2 at	1 20000104	L	Licitaspair o

<u> </u>	204028	DEEGOOO	T	Leutativo III A class II acces protein I
51	201038 s at	BE560202		putative HLA class II assoc protein I
52	201051_at	BE560202	100104	putative HLA class II assoc protein I
53	201105_at	NM 002305	LGALS1	lectin, galactoside-binding, soluble, 1
54	201106_at	NM_002085	GPX4	glutathione peroxidase 4
55	201188_s_at	D26351	ITPR3	type 3 inositol 1,4,5-trisphosphate
56	201190	NIM OCCOSA	ITPR3	receptor
56	201189 s at 201195 s at	NM_002224 AB018009	IIPRO	I type amine said transporter 4
57	201195 s at	NM 002592	PCNA	L-type amino acid transporter 1 proliferating cell nuclear antigen
58		NM 002654	PKM2	pyruvate kinase, muscle
59 60	201251_at 201252_at	NM 006503	PSMC4	proteasome 26S subunit, ATPase, 4
61	201252_at	NM 016732	RALY	RNA-binding protein transcript var 1
62	201271 s_at	NM 001067	10461	topoisomerase (DNA) II alpha
63	201313 at	NM 001975	ENO2	enolase 2
	201315_at	NM 024551	FLJ21432	hypothetical protein FLJ21432
64 65	201346_at	NM 000876	IGF2R	insulin-like growth factor 2 receptor
66	201416 at	NM 003107	101 211	SRY-box 4
67	201410_at	NM 003107		SRY-box 4
68	201477_at	NM_004832	GSTTLp28	glutathione-S-transferase like;
	201710_at	14101_007002	J 3011 Ep20	glutathione transferase omega
69	201474_s_at	NM 002204	ITGA3	integrin, alpha 3 transcript variant a
70	201485_s_at	BC004892	RCN2	reticulocalbin 2, EF-hand calcium
				binding domain
71	201486 at	NM 002902	RCN2	
72	201536 at	AL048503		DKFZp586M1524
73	201614_s_at	NM_003707	RUVBL1	RuvB (E coli homolog)-like 1
74	201660_at		FACL3	fatty-acid-Coenzyme A ligase, long-
				chain 3
75	201661_s_at	NM_004457	FACL3	
76	201662_s_at	D89053		Acyl-CoA synthetase 3
77	201670_s_at	M68956	MARCKS 80K- L	myristoylated alanine-rich C-kinase substrate
78	201714_at	NM_001070	TUBG1	tubulin, gamma 1
79	201765 s_at	AL523158		hexosaminidase A
80	201792_at	NM_001129	AEBP1	AE-binding protein 1
81	201804_x_at	NM_001281	CKAP1	cytoskeleton-associated protein 1
82	201819_at	NM_005505	CD36L1	CD36 antigen-like 1
83	201850_at	NM_001747	CAPG	capping protein gelsolin-like
84	201880_at	NM_005744		ariadne (Drosophila) homolog,
				ubiquitin-conjugating enzyme E2-
L		75040275	54554	binding protein, 1
85	201910_at	BF213279	FARP1	RhoGEF & pleckstrin domain 1
86	201911 s at	NM_005766	FARP1	de de la companya de
87	201931_at	NM_000126	ETFA	electron-transfer-flavoprotein, α polypeptide
88	201954_at	NM_005720	ARPC1B	actin related protein 23 com, sub 1A
89	201976_s_at	NM_012334	MYO10	myosin X
90	202069_s_at	Al826060	IDH3A	isocitrate dehydrogenase 3 alpha
91	202070_s_at	NM_005530	IDH3A	
92	202111_at	NM_003040	SLC4A2	solute carrier fam 4 anion exchanger
				mem 2
93	202154_x_at	NM_006086	TUBB4	tubulin, beta, 4
94	202185_at	NM_001084	PLOD3	procollagen-lysine, 2-oxoglutarate 5-
				dioxygenase

	000400 -1	1 104 044000	1444000	
95	202188_at	NM_014669	KIAA0095	KIAA0095 gene product
96	202219_at	NM_005629	SLC6A8	solute carrier family 6, member 8
97	202224_at	NM_016823		v-crk avian sarcoma virus CT10
			 	oncogene homolog
98	202225_at	NM_016823		v-crk avian sarcoma virus CT10
				oncogene homolog
99	202260_s_at	NM_003165	STXBP1	syntaxin binding protein 1
100	202295_s_at	NM_004390	CTSH	cathepsin H
101	202329_at	NM_004383	CSK	c-src tyrosine kinase
102	202367_at	NM_001913	CUTL1	cut (Drosophila)-like 1
103	202370_s_at	NM_001755	CBFB	core-binding factor, β sub trans var 2
104	202478_at	NM_021643	GS3955	GS3955 protein
105	202503_s_at	NM_014736	KIAA0101	KIAA0101 gene product
106	202589_at	NM_001071	TYMS	thymidylate synthetase
107	202603_at	N51370		disintegrin and metalloproteinase
				domain 10
108	202705_at	NM_004701	CCNB2	cyclin B2
109	202737_s_at	NM_012321	LSM4	U6 snRNA-associated Sm-like protein
110	202779_s_at	NM_014501	E2-EPF	ubiquitin carrier protein
111	202785_at	NM_005001	NDUFA7	NADH dehydrogenase 1 α
				subcomplex, 7
112	202862_at	NM_000137	FAH	fumarylacetoacetate
113	202898_at	NM_014654	KIAA0468	KIAA0468 gene product
114	202954_at	NM_007019	UBCH10	ubiquitin carrier protein E2-C
115	202958_at	NM_002833	PTPN9	protein tyrosine phosphatase, non-
	_	_		receptor type 9
116	- 202961_s_at	NM_004889	ATP5J2	ATP synthase, H+ transporting,
		_		mitochondrial F0 complex, subunit f,
				isoform 2
117	202986_at	NM_014862	KIAA0307	KIAA0307 gene product
118	203011_at	NM_005536	IMPA1	inositol(myo)-1(or 4)-
				monophosphatase 1
119	203022_at	NM_006397	RNASEHI	ribonuclease HI, large subunit
120	203069_at	NM_014849	KIAA0736	KIAA0736 gene product
121	203071_at	NM_004636	SEMA3B	sema domain, Ig domain, short basic
	<u> </u>			domain, secreted, 3B
122	203094_at	NM_014628	KIAA0110	gene predicted from cDNA
123	203145_at	NM_006461	DEEPEST	mitotic spindle coiled-coil related
124	203167_at	NM_003255	TIMP2	tissue inhibitor of metalloproteinase 2
125	203217_s_at	NM_003896	SIAT9	sialyltransferase 9
126	203234_at	NM_003364	UP	uridine phosphorylase
127	203256_at	NM_001793	CDH3	cadherin 3, type 1, P-cadherin
				(placental)
128	203262_s_at	NM_004699	DXS9928E	chromosome X 9928 expressed seq
129	203300_x_at	NM_003916	AP1S2	adaptor-related protein complex 1,
				sigma 2 subunit
130	203315_at	BC000103		NCK adaptor protein 2,
131	203366_at	NM_002693	POLG	polymerase (DNA directed), gamma
132	203396_at	NM_002789	PSMA4	proteasome subunit, α type, 4
133	203452_at	NM_012200	B3GAT3	beta-1,3-glucuronyltransferase 3
134	203456_at	NM_007213	JM4	JM4 protein
135	203502 at	NM_001724	BPGM	2,3-bisphosphoglycerate mutase
136	203518 at	NM_000081	CHS1	Chediak-Higashi syndrome 1
137	203554_x_at	NM_004219	PTTG1	pituitary tumor-transforming 1

	100055	104 00000	DODD	
138	203557_s_at	NM_000281	PCBD	6-pyruvoyl-tetrahydropterin
				synthasedimerization cofactor of
400	000570 -4	NIN 005570	1.00//	hepatocyte nuclear factor 1 alpha
139	203570_at	NM_005576	LOXL1	lysyl oxidase-like 1
140	203590_a t	NM_006141	DNCLI2	dynein, cytoplasmic, light intermediate
144	000040 -4	NIN4 000404	EDE	polypeptide 2
141	203643_at	NM_006494	ERF	Ets2 repressor factor
142	203663 s_at	NM_004255	COX5A	cytochrome c oxidase subunit Va
143	203668_at	NM_006715	MAN2C1	mannosidase, α, class 2C, mem 1
144	203693 s at	NM_001949	E2F3	E2F transcription factor 3
145	203695 s at	NM_004403	DFNA5	deafness, autosomal dominant 5
146	203723_at	NM_002221	ITPKB	inositol 1,4,5-trisphosphate 3-kinase B
147	203729_at	NM_001425	EMP3	epithelial membrane protein 3
148	203730_s_at	BF196931	ZFP95	zinc finger protein homologous to
140	0007041	NNA 044500	75505	Zfp95 in mouse
149	203731 s at	NM_014569	ZFP95	1 0 1 0
150	203775_at	NM_014251	SLC25A13	solute carrier family 25, member 13
151	203827_at	NM_017983	FLJ10055	hypothetical protein FLJ10055
152	203878_s_at	NM_005940	MMP11	matrix metalloproteinase 11
153	204014_at	NM_001394	DUSP4	dual specificity phosphatase 4
154	204015 s at	BC002671	DUSP4	
155	204033_at	NM_004237	TRIP13	thyrold hormone receptor interactor 13
156	204092 s_at	NM_003600	STK15	serinethreonine kinase 15
157	204099_at	NM_003078	SMARCD3	SWISNF related, matrix associated,
Ì		İ	1	actin dependent regulator of
450	004470	NM 001827	CKS2	chromatin, subfamily d, member 3
158	204170_s_at		RUNX3	CDC28 protein kinase 2
159	204197_s_at 204198 s at	NM_004350 AA541630	RUNX3	runt-related transcription factor 3
160	204198_s_at	NM 017604	KIAA1023	KIAA1023 protein
161 162	204202_at	NM 006347	USA-CYP	cyclophilin
163	204226_at	NM 006716	ASK	activator of S phase kinase
164	204244_s_at	NM_004935	CDK5	cyclin-dependent kinase 5
165	204247 s at	M68520	CDRO	cdc2-related protein kinase
166	204252_at	NM 000447	PSEN2	presenilin 2 transcript variant 1
167	204202 s at	NM 013255	MKLN1	muskelin 1, intracellular mediator
107	204425_at	14141_013233	WINCENT	containing kelch motifs
168	204436 at	NM 025201	PP1628	hypothetical protein PP1628
169	204458 at	AL110209	111020	DKFZp564A0122
170	204467_s_at	NM 000345	SNCA	synuclein, α transcript var NACP140
171	204584_at	Al653981	L1CAM	L1 cell adhesion molecule, MASA
'''	20 100u.	,		transcript var 1
172	204585 s at	NM_000425	L1CAM	
173	204647 at	NM 004838	HOMER-3	Homer, neuronal imm early gene, 3
174	204654_s_at	NM 003220	TFAP2A	transcription factor AP-2 alpha
175	204709_s_at	NM_004856	KNSL5	kinesin-like 5
176	204778_x_at	AW102783	HOXB7	homeo box B7
177	204779 s at	NM_004502	HOXB7	
178	204857_at	NM_003550	MAD1L1	MAD1-like 1
179	204932_at	BF433902		TNF receptor superfam, mem 11b
180	204973 at	NM_000166	GJB1	gap junction protein, beta 1, 32kD
181	204995 at	AL567411		cyclin-dependent kinase 5, regulatory
'-'				sub 1 (p35)
182	205051 s at	NM_000222	KIT	v-kit Hardy-Zuckerman 4 feline
			· · · · · · · · · · · · · · · · · · ·	1

Sarcoma viral oncogene homolog Sarcoma viral oncogene homolog ABCD1 ATP-binding cassetle, sub-family D (ALD), mem 1		- 			
CALD), mem 1		 		 	
184	183	205142_x_at	NM_000033	ABCD1	
185					
187					
Phosphatase, type II, 105kD					
188	186	205376_at	NM_003866	INPP4B	
188					
Short cytoplasmic domain 5A mitogen-activated protein kinase kinase kinase 12	187	205405_at	NM_003966	SEMA5A	
188				ŀ	
Rinase kinase 12 Rinase kinase 12 Rinase kinase 12 Rinase 22 Rinase 22 Rinase 22 Rinase 23 Rinase 24 Rinase 24 Rinase 25 Rinase 25 Rinase 26 Rinase 26 Rinase 26 Rinase 27 Rinase 26 Rinase 27 Rinase 26 Rinase 27 Rinase 26 Rinase 27 Rinase	100	 			
189	188	205447_s_at	BE222201		
90	400	005450 -1	D0004070		
191 205591 at				1104.0	
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233 208744 x at BG403660 heat shock 105kD 234 208837 at BC000027 integral type I protein 235 208916 at AF105230 SLC1A5 neutral amino acid transporter 236 208928 at AF258341 NADPH-cytochrome P450 reductase 237 208956 x at U62891 DUT deoxyuridine triphosphatase 238 208974 x at BC003572 karyopherin (importin) beta 1 239 208975 s at L38951 Importin beta subunit 240 209015 s at BC002446 MRJ gene for a member of DNAJ fair 241 209036 s at BC001917 malate dehydrogenase 2, NAD 242 209053 s at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072 at M13577 MBP myelin basic protein 244 209079_x at AF152318 PCDH-gamma- protocadherin gamma A1 245 209081_s at BC000576 quinoid dihydropteridine reductase 247 209132_s at BE313890 hypothetical protein FLJ20452 <tr< td=""></tr<>
234 208837 at BC000027 SLC1A5 Integral type protein 235 208916 at AF105230 SLC1A5 NADPH-cytochrome P450 reductase 237 208956 x_at U62891 DUT deoxyuridine triphosphatase 238 208974 x_at BC003572 Raryopherin (importin) beta 1 239 208975 s_at L38951 Importin beta subunit 240 209015 s_at BC002446 MRJ gene for a member of DNAJ far 241 209036 s_at BC001917 Importin beta subunit 242 209053 s_at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072 at M13577 MBP myelin basic protein 244 209079_x_at AF152318 PCDH-gamma-
235 208916_at AF105230 SLC1A5 neutral amino acid transporter 236 208928_at AF258341 NADPH-cytochrome P450 reductase 237 208956_x_at U62891 DUT deoxyuridine triphosphatase 238 208974_x_at BC003572 karyopherin (importin) beta 1 239 208975_s_at L38951 importin beta subunit 240 209015_s_at BC002446 MRJ gene for a member of DNAJ fair 241 209036_s_at BC001917 malate dehydrogenase 2, NAD 242 209053_s_at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072_at M13577 MBP myelin basic protein 244 209079_x_at AF152318 PCDH-gamma-A1 PCDH-gamma-A1 PCDH-gamma-A1 PCDH-gamma-A1 245 209123_at BC000576 quinoid dihydropteridine reductase 247 209132_s_at BE313890 hypothetical protein FLJ20452 248 209172_s_at J30872 mitosin 249 209197_at AA626780 KIAA0080 protein 250 209198_s_at BC001661 ATP-binding cassette, sub-fam F mem 2 252 209254_at AF277177 KIAA0265_protein 253 209255_at AF277177 KIAA0265_protein 254 209256_s_at AF277177 PNAS-119 255 209283_at AL561930 phosphatidylinositol_4-kinase_type_II
236 208928_at
237 208956_x_at U62891 DUT deoxyuridine triphosphatase 238 208974_x_at BC003572 karyopherin (importin) beta 1 239 208975_s_at L38951 importin beta subunit 240 209015_s_at BC002446 MRJ gene for a member of DNAJ far 241 209036_s_at BC001917 malate dehydrogenase 2, NAD 242 209053_s_at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072_at M13577 MBP myelin basic protein 244 209079_x_at AF152318 PCDH-gamma-A1 protocadherin gamma A1 245 209081_s_at NM_030582 COL18A1 collagen, type XVIII, alpha 1 246 209123_at BC000576 quinoid dihydropteridine reductase 247 209132_s_at BE313890 hypothetical protein FLJ20452 248 209172_s_at Jake 20918 Jake 20918 Jake 20918 250 209188_s_at BC001661 ATP-binding cassette, sub-fam F 251 209254_at AF277177 KIAA0265 protein </td
238 208974 x at 209075 s at L38951 karyopherin (importin) beta 1 239 208975 s at L38951 importin beta subunit 240 209015 s at BC002446 MRJ gene for a member of DNAJ fair and the provided subunit 241 209036 s at BC001917 malate dehydrogenase 2, NAD 242 209053 s at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072 at M13577 MBP myelin basic protein 244 209079_x_at AF152318 PCDH-gamma-A1 protocadherin gamma A1 245 209081 s at BC000576 Quinoid dihydropteridine reductase 247 209123 at BE313890 hypothetical protein FLJ20452 248 209172 s at U30872 mitosin 249 209197 at AA626780 KIAA0080 protein 250 209198 s at BC001661 ATP-binding cassette, sub-fam F mem 2 252 209254 at AF277177 KIAA0265 protein 253 209255 at AF277177 KIAA0265 protein 254 209256 s at AF277177 PNAS-119 255 209283 at AF007162 unknown mRNA 256 209345 s at AL561930
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239 208975_s_at L38951 importin beta subunit
241 209036 s at BC001917 malate dehydrogenase 2, NAD 242 209053 s at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072 at M13577 MBP myelin basic protein 244 209079 x at AF152318 PCDH-gamma-A1 245 209081 s at NM_030582 COL18A1 collagen, type XVIII, alpha 1 246 209123 at BC000576 quinoid dihydropteridine reductase 247 209132 s_at BE313890 hypothetical protein FLJ20452 248 209172 s_at U30872 mitosin 249 209197 at AA626780 KIAA0080 protein 250 209198 s_at BC001661 Similar to synaptotagmin 11 251 209247_s_at BC001661 ATP-binding cassette, sub-fam F mem 2 252 209254_at AF277177 KIAA0265 protein 253 209255_at AF277177 PNAS-119 255 209283_at AF007162 unknown mRNA 256 209345_s_at AL561930 phosphatidylinositol 4-kinas
241 209036 s at BC001917 malate dehydrogenase 2, NAD 242 209053 s at AF083389 Wolf-Hirschhorn syn candidate 1 243 209072 at M13577 MBP myelin basic protein 244 209079 x at AF152318 PCDH-gamma-A1 245 209081 s at NM_030582 COL18A1 collagen, type XVIII, alpha 1 246 209123 at BC000576 quinoid dihydropteridine reductase 247 209132 s_at BE313890 hypothetical protein FLJ20452 248 209172 s_at U30872 mitosin 249 209197 at AA626780 KIAA0080 protein 250 209198 s_at BC001661 Similar to synaptotagmin 11 251 209247_s_at BC001661 ATP-binding cassette, sub-fam F mem 2 252 209254_at AF277177 KIAA0265 protein 253 209255_at AF277177 PNAS-119 255 209283_at AF007162 unknown mRNA 256 209345_s_at AL561930 phosphatidylinositol 4-kinas
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243 209072_at M13577 MBP myelin basic protein 244 209079_x_at AF152318 PCDH-gamma-A1 245 209081_s_at NM_030582 COL18A1 collagen, type XVIII, alpha 1 246 209123_at BC000576 quinoid dihydropteridine reductase 247 209132_s_at BE313890 hypothetical protein FLJ20452 248 209172_s_at U30872 mitosin 249 209197_at AA626780 KIAA0080 protein 250 209198_s_at BC004291 Similar to synaptotagmin 11 251 209247_s_at BC001661 ATP-binding cassette, sub-fam F mem 2 252 209254_at AF277177 KIAA0265 protein 253 209255_at AF277177 RIAA0265 protein 254 209256_s_at AF277177 PNAS-119 255 209283_at AF007162 unknown mRNA 256 209345_s_at AL561930 phosphatidylinositol 4-kinase type II
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249 209197 at AA626780 KIAA0080 protein 250 209198 s at BC004291 Similar to synaptotagmin 11 251 209247 s at BC001661 ATP-binding cassette, sub-fam F mem 2 252 209254 at AF277177 KIAA0265 protein 253 209255 at AF277177 KIAA0265 protein 254 209256 s at AF277177 PNAS-119 255 209283 at AF007162 unknown mRNA 256 209345 s at AL561930 phosphatidylinositol 4-kinase type II
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255 209283_at AF007162 unknown mRNA 256 209345_s_at AL561930 phosphatidylinositol 4-kinase type II
256 209345_s_at AL561930 phosphatidylinositol 4-kinase type II
truncated suppressin
258 209515 s at U38654 Rab27a
259 209773_s_at BC001886 ribonucleotide reductase M2 polyper
260 209825_s_at BC002906 Sim to uridine monophosphate kinas
261 209827_s_at NM_004513 IL16 interleukin 16
262 209828_s_at M90391 putative IL-16 protein precursor
263 209848_s_at U01874 me20m
264 209875_s_at M83248 nephropontin
265 209932_s_at U90223 deoxyuridine triphosphate
nucleotidohydrolase precursor
266 210052_s_at AF098158 restricted expressed proliferation
associated protein 100
267 210073_at L32867 alpha 2,8-sialyltransferase
268 210111 s_at AF277175 PNAS-138 269 210127 at BC002510 small GTPase RAB6B

270	210854 x at	U17986		GABAnoradrenaline transporter
271	210926 at	AY014272	FKSG30	FKSG30
272	210948 s at	AF294627	LEF1	lymphoid enhancer factor 1 isoform
273	210951 x at	AF125393		Rab27 isoform
274	211013 x at	AF230411		tripartite motif protein TRIM19 lambda
275	211052 s at	BC006364		clone MGC:12705,
276	211066 x at	BC006439		Similar to protocadherin gamma
- ' -				subfamily A, 5
277	211373 s at	U34349	AD3LPAD5	seven trans-membrane domain
278	211564 s at	BC003096		Sim to LIM domain protein
279	211752 s_at	BC005954		clone MGC:14592
280	211759 x at	BC005969		clone MGC:14625
281	211833 s at	U19599		BAX delta
282	212000_at	AB002363	***	KIAA0365 gene product
283	212070_at	AL554008	<u> </u>	G protein-coupled receptor 56
284	212081_x_at	AF129756	MSH55	MSH55
285	212119 at	BF348067		phosphatidylinositol glycan, class F
286	212178_s_at	AK022555		FLJ12493 fis
287	212193 s_at	BE881529		KIAA0731 protein
288	212247_at	AW008531		KIAA0225 protein
289	212285 s at	AF016903		IMAGE:3506210
290	212312 at	AL117381		clone RP5-857M17 on chrom 20
291	212338 at	AA621962		KIAA0727 protein
292	212402_at	BE895685		KIAA0853 protein
293	212472 at	BE965029		FLJ22463 fis
294	212473 s_at	BE965029		FLJ22463 fis
295	212512 s at	AA551784	•	coactivator-associated arginine
				methyltransferase-1
296	212520 s at	Al684141		SWISNF related
297	212552 at	BE617588		hippocalcin-like
298	212715_s_at	AB020626		KIAA0819 protein
299	212739_s_at	AL523860		non-metastatic cells 4
300	212744_at	Al813772		clone HQ0692
301	212745_s_at	Al813772		clone HQ0692
302	212793_at	BF513244		KIAA0381 protein
303	212796_s_at	BF195608		KIAA1055 protein
304	213002_at	BF347326	MARCKS, 80K-	myristoylated alanine-rich protein
			L	kinase C substrate
305	213007_at	BG478677		polymerase (DNA directed), gamma
306	213008_at	BG478677		polymerase (DNA directed), gamma
307	213096_at	T51252		KIAA0481 gene product
308	213131_at	R38389		olfactomedin related ER localized
309	213169_at	BG109855		clone TUA8 Cri-du-chat region
310	213215_at	Al910895	<u> </u>	clone EUROIMAGE 42138
311	213217_at	AU149572		adenylate cyclase 2(brain)
312	213241_at	AF035307		clone 23785
313	213274 s at	BE875786		cathepsin B
314	213275 x at	BE875786		cathepsin B
315	213330 s at	BE886580		stress-induced-phosphoprotein 1
316	213333_at	AL520774		malate dehydrogenase 2, NAD
317	213338_at	BF062629		DKFZP586E1621 protein
318	213392_at	AW070229		G protein-coupled receptor, fam C, group 5, mem B
319	213474 at	AI890903		ESTs
				*

321 213573 at AA861808 karyopherin (importin) beta 1 322 213587 s at Al884867 ribosomal protein L28 323 213638 at AW054711 PAC 257A7 on chromosome 6p24 324 213670 x at Al768378 KIAA0618 gene product SWISNF related, matrix associated actin dependent regulator of chromatin, subfam a, member 4 filamin A, alpha 326 213746 s at AW051856 filamin A, alpha 328 213836 s at AW052084 KIAA1001 protein 329 213980 at T87225 CLONE=IMAGE:22392 330 214023 x at AL533838 tubulin, beta polypeptide 331 214068 at AF070610 clone 24505 332 214104 at Al703188 G-protein coupled receptor 414081 x at Al738662 homeo box H89 335 214614 at Al738662 homeo box H89 336 214682 at AA295257 neuropilin 2 myosin IB 339 214708 at BG484314 syntrophin, beta 1 214714 at AK022360 FLJ12298 fis AK022360 FLJ12298 fis AB41414 AB417834 AB41414 AK022360 FLJ12298 fis AB41784 AB41841 AB5484 AB618414 AK022360 FLJ12298 fis AB41841 AB611541 AB61841 AB611541 AB61841 AB611541 AB61841 AB61854 AB61855		γ	1		
323	320	213496_at	AW592563		KIAA0455 gene product
233 213638 at AW054711 PAC 257A7 on chromosome 6p24					
325					
325					
actin dependent regulator of chromatin, subfam a, member 4					
Chromatin, subfam a, member 4	325	213720_s_at	Al831675		
326	1				actin dependent regulator of
327 213836 s at					chromatin, subfam a, member 4
328 213995 at BF445047 spithelial membrane protein 1 329 213960 at T87225 CLONE=IMAGE:22392 330 214023 x at AL533838 tubulin, beta polypeptide 331 214068 at AF070610 clone 24505 Clone 24505 332 214104 at Al703188 G-protein coupled receptor 333 214201 x at AA742237 HLA-B associated transcript-2 334 214581 x at BE568134 death receptor 6 335 214614 at Al738662 homeo box HB9 336 214632 at AA295257 neuropilin 2 337 214656 x at BE790157 myosin IB 338 214687 x at AK026577 FLJ22924 fis 339 214708 at BE407516 cyclin B1 341 214714 at AK022360 FLJ12298 fis 342 214717 at AL137534 DKFZp434H1419 343 214752 x at AB025550 filamin A, alpha 344 214778 at AB011541 MEGF8 346 214893 x at Al421964 Al29647 Al29647 Al29647 Al29647 Al29647 Al29647 Al29648 Al29647 Al29647 Al29648 Al29647 Al29647 Al29648 Al29647 Al29648 Al29647 Al29648 Al29647 Al29648 Al29647 Al29648 Al29648 Al29648 Al29648 Al29648 Al29667 Al29648 Al29668 Al29674 Al29668 Al29668 Al29678 Al29678 Al29668 Al29678 Al29668 Al29678 Al29668 Al29678 Al29668 Al29678 Al296					filamin A, alpha
329 213960 at	327	213836_s_at	AW052084		KIAA1001 protein
329	328	213895_at	BF445047		epithelial membrane protein 1
331 214068 at AF070610 Clone 24505	329	213960 at	T87225		
331 214088 at AF070610 Clone 24505	330	214023 x at	AL533838		tubulin, beta polypeptide
332 214104 at Al703188 G-protein coupled receptor					
333					
334					
335		214581 x at			
336 214632 at AA295257 neuropilin 2					
337 214656 x at BE790157 myosin IB 338 214687 x at AK026577 FLJ22924 fis 339 214708 at BG484314 syntrophin, beta 1 340 214710 s at BE407516 cyclin B1 341 214714 at AK022360 FLJ12298 fis 342 214717 at AL137534 DKFZp434H1419 343 214752 x at AI625550 filamin A, alpha 344 214778 at AB011541 MEGF8 345 214841 at AF070524 clone 24453 346 214893_x_at AI21964 hyperpolarization activated cyclic nucleotide-gated potassium channel 347 214896 at AL109671 EUROIMAGE 29222 348 215025 at S76476 trkC {alternatively spliced} 349 215115 x at AI613045 ets variant gene 6 (TEL oncogene) 350 215126 at AL109716 EUROIMAGE 208948 351 215155 at J04178 HEXA abnormal β-hexosaminidase α chair 352 215311 at AL109696 EUROIMAGE 21920 354 215836 s at AK026188 FLJ22535 fis 355 216194_s_at AD001527 DNA from chrom 19-cosmid f24590 356 216973_s_at S76475 trkC neurotrophic tyrosine kinase, receptor, type 3 358 217104_at AL109714 EUROIMAGE 327506 359 217226 s at M95929 PHOX1 Paired mesoderm homeo box 1 360 217297_s_at AF143684 MYO9b unconventional myosin IXb 361 217377_x_at AF041811 ETV6-NTRK3 ETS related protein-growth factor receptor tyrosine kinase fusion					
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339					
340 214710_s_at BE407516 Cyclin B1 341 214714_at AK022360 FLJ12298 fis 342 214717_at AL137534 DKFZp434H1419 343 214752_x_at Al625550 filamin A, alpha 344 214778_at AB011541 MEGF8 345 214841_at AF070524 Clone 24453 346 214893_x_at Al421964 hyperpolarization activated cyclic nucleotide-gated potassium channe 347 214896_at AL109671 EUROIMAGE 29222 348 215025_at S76476 trkC {alternatively spliced} 349 215115_x_at Al613045 ets variant gene 6 (TEL oncogene) 350 215126_at AL109716 EUROIMAGE 208948 351 215155_at J04178 HEXA abnormal β-hexosaminidase α chair 352 215311_at AL109696 EUROIMAGE 21920 353 215812_s_at U41163 SLC6A10 creatine transporter 354 215836_s_at AK026188 FLJ22535 fis 355 216194_s_at AD001527 DNA from chrom 19-cosmid f24590 356 216973_s_at S49765 homeo box B7 357 217033_x_at S76475 trkC neurotrophic tyrosine kinase, receptor, type 3 358 217104_at AL109714 EUROIMAGE 327506 359 217226_s_at M95929 PHOX1 Paired mesoderm homeo box 1 360 217297_s_at AF143684 MYO9b unconventional myosin IXb 361 217377_x_at AF041811 ETV6-NTRK3 ETS related protein-growth factor receptor tyrosine kinase fusion					
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343					
344 214778_at AB011541 MEGF8 clone 24453 345 214841_at AF070524 clone 24453 hyperpolarization activated cyclic nucleotide-gated potassium channe 347 214896_at AL109671 EUROIMAGE 29222 348 215025_at S76476 trkC {alternatively spliced} 349 215115_x_at Al613045 ets variant gene 6 (TEL oncogene) 350 215126_at AL109716 EUROIMAGE 208948 351 215155_at J04178 HEXA abnormal β-hexosaminidase α chair 352 215311_at AL109696 EUROIMAGE 21920 353 215812_s_at U41163 SLC6A10 creatine transporter 354 215836_s_at AK026188 FLJ22535 fis 355 216194_s_at AD001527 DNA from chrom 19-cosmid f24590 containing CAPNS and POL2RI 356 216973_s_at S76475 trkC neurotrophic tyrosine kinase, receptor, type 3 358 217104_at AL109714 EUROIMAGE 327506 359 217226_s_at M95929 PHOX1 Paired mesoderm homeo box 1 360 217297_s_at AF041811 ETV6-NTRK3 ETS related protein-growth factor receptor tyrosine kinase fusion					
345					
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nucleotide-gated potassium channe 347					
347	340	214093_X_at	A142 1304		nucleatide geted netersium channel 2
348	247	214906 ot	AL 100671	ļ	FUDOMACE 20222
349				· · · · · · · · · · · · · · · · · · ·	
350 215126 at AL109716 EUROIMAGE 208948 351 215155 at J04178 HEXA abnormal β-hexosaminidase α chair 352 215311 at AL109696 EUROIMAGE 21920 353 215812 s at U41163 SLC6A10 creatine transporter 354 215836 s at AK026188 FLJ22535 fis 355 216194 s at AD001527 DNA from chrom 19-cosmid f24590				·. · · · · · · · · · · · · · · · · · ·	
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363 217624_at AA464753 ESTs	363				
364 217799 x at NM_003344 UBE2H ubiquitin-conjugating enzyme E2H	364		NM_003344		ubiquitin-conjugating enzyme E2H
365 217827 s_at NM_016630 ACP33 acid cluster protein 33	365				acid cluster protein 33
366 217867_x_at NM_012105 BACE2 beta-site APP-cleaving enzyme 2			NM_012105	BACE2	

368 217891 at NM 022744 FLJ13888 hypothetical protein FLJ13888 1809 218090 s at NM 003981 PRC1 protein regulator of cytokinesis 1 370 218030 at NM 014030 GIT1 G protein-coupled receptor kinase-interactor 1 G protein-coupled receptor kinase-interactor 1 218074 at NM 016062 LOC51647 CGI-128 protein ST21413 s at NM 024531 FLJ11856 hypothetical protein FLJ20561 hypothetical protein FLJ10333 hypothetical protein FLJ10333 hypothetical protein FLJ10333 hypothetical protein FLJ10334 hypothetical protein FLJ10335 hypothetical protein FLJ10335 hypothetical protein FLJ10335 hypothetical protein FLJ10337 hypothetical protein FLJ10520 hypothetical protein FLJ10533 hypothetical protein FLJ10430 hypothetical protein FLJ10430 hypothetical protein FLJ10430 hypo	367	217874_at	NM 003849	SUCLG1	succinate-CoA ligase, GDP-forming.
368 217891 at NM 022744 FLJ13868 hypothetical protein FLJ13868 369 218009 s at NM 003981 PRC1 protein regulator of cytokinesis 1 370 218074 at NM 014030 GIT1 G protein-coupled receptor kinase-interactor 1 371 218074 at NM 016062 LOC51647 CG-128 protein 372 218153 at NM 005697 SCAMP2 secretory carrier membrane protein 2 373 218151 x at NM 018200 HMG20A high-mobility group 20A 375 218153 at NM 017882 FLJ20561 hypothetical protein FLJ10633 376 218175 at NM 021495 FLJ20561 hypothetical protein FLJ20561 377 218330 s at NM 017875 FLJ10033 hypothetical protein FLJ10033 378 218349 s at NM 024958 FLJ23329 hypothetical protein FLJ10036 379 218359 at NM 024958 FLJ23329 hypothetical protein FLJ10333 380 218542 at NM 021042 FLJ23329 hypothetical protein FLJ10540 381	307	21/0/4_at	14141_003049	SUCLGI	J 9 1
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389 218839_at	387	218786_at		TU12B1-TY	TU12B1-TY protein
YRPW motif 1			NM_018215	FLJ10781	hypothetical protein FLJ10781
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393 218952_at NM_013271 SAAS granin-like neuroendocrine peptide precursor 394 218956_s_at NM_015545 KIAA0632 KIAA0632 protein 395 218980_at NM_025135 KIAA1695 hypothetical protein FLJ22297 396 218996_at NM_025135 KIAA1695 hypothetical protein FLJ22297 397 219011_at NM_020904 PLEKHA4 pleckstrin homology domain-containing, family A member 4 398 219039_at NM_017789 FLJ20369 hypothetical protein FLJ20369 399 219040_at NM_024535 FLJ22021 hypothetical protein FLJ20201 400 219041_s_at NM_014374 AP4 zinc finger protein 401 219051_x_at NM_024042 MGC2601 hypothetical protein MGC2601 402 219066_at NM_021823 MDS018 hypothetical protein MDS018 403 219066_at NM_021823 MDS018 hypothetical protein MDS018 404 219148_at NM_018492 TOPK PDZ-binding kinase; T-cell originated protein kinase 405 219152_at NM_017854 FLJ20512 hypothetical protein FLJ20512 407 219361_s_at NM_022767 FLJ12484 hypothetical protein FLJ20512 408 219408_at NM_014055 CDV-1 CDV-1 protein 409 219408_at NM_014055 CDV-1 CDV-1 protein FLJ10640 410 219478_at NM_021197 WFDC1 WAP four-disulfide core domain 1 411 219491_at NM_024036 MGC3103 hypothetical protein MGC3103 412 219522_at NM_014344 FJX1 putative secreted ligand homologous to fjx1					hypothetical protein FLJ10430
Description				FLJ12538	
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407 219361 s at NM 022767 FLJ12484 hypothetical protein FLJ12484 408 219372 at NM 014055 CDV-1 CDV-1 protein 409 219408 at NM 019023 FLJ10640 hypothetical protein FLJ10640 410 219478 at NM 021197 WFDC1 WAP four-disulfide core domain 1 411 219491 at NM 024036 MGC3103 hypothetical protein MGC3103 412 219522 at NM_014344 FJX1 putative secreted ligand homologous to fjx1					
408 219372 at NM 014055 CDV-1 CDV-1 protein 409 219408 at NM 019023 FLJ10640 hypothetical protein FLJ10640 410 219478 at NM 021197 WFDC1 WAP four-disulfide core domain 1 411 219491 at NM 024036 MGC3103 hypothetical protein MGC3103 412 219522_at NM_014344 FJX1 putative secreted ligand homologous to fjx1					
409 219408 at NM 019023 FLJ10640 hypothetical protein FLJ10640 410 219478 at NM 021197 WFDC1 WAP four-disulfide core domain 1 411 219491 at NM 024036 MGC3103 hypothetical protein MGC3103 412 219522 at NM_014344 FJX1 putative secreted ligand homologous to fjx1					
410219478_atNM_021197WFDC1WAP four-disulfide core domain 1411219491_atNM_024036MGC3103hypothetical protein MGC3103412219522_atNM_014344FJX1putative secreted ligand homologous to fjx1					
411219491_atNM_024036MGC3103hypothetical protein MGC3103412219522_atNM_014344FJX1putative secreted ligand homologous to fjx1					hypothetical protein FLJ10640
412 219522_at NM_014344 FJX1 putative secreted ligand homologous to fjx1					
to fjx1		219491_at		MGC3103	
	412	219522_at	NM_014344	FJX1	putative secreted ligand homologous
413 219537 x at NM_016941 DLL3 Delta (Drosophila)-like 3					
	413	219537_x_at	NM_016941	DLL3	Delta (Drosophila)-like 3

				
414	219555_s_at	NM_018455	BM039	uncharacterized bone marrow protein BM039
415	219578_s_at	NM_030594	FLJ13203	hypothetical protein FLJ13203
416	219634_at	NM_018413	C4ST	chondroitin 4-sulfotransferase
417	219637_at	NM_025139	FLJ12584	hypothetical protein FLJ12584
418	219703 at	NM 018365	FLJ11222	hypothetical protein FLJ11222
419	219742 at	NM 030567	MGC10772	hypothetical protein MGC10772
420	219895 at	NM 017938	FLJ20716	hypothetical protein FLJ20716
421	219933 at	NM 016066	LOC51022	CGI-133 protein
422	220116_at	NM_021614	KCNN2	potassium intermediatesmall
				conductance calcium-activated
		1		channel, subfamily N, member 2
423	220155 s at	NM_023924	FLJ13441	hypothetical protein FLJ13441
424	220178 at	NM 021731	PP3501	hypothetical protein PP3501
425	220454 s_at	NM 020796	SEMA6A	sema domain, transmembrane
,0		''''_'		domain and cytoplasmic domain, 6A
426	220864_s_at	NM 015965	LOC51079	CGI-39 protein; cell death-regulatory
		10.10000		protein GRIM19
427	220948_s_at	NM 000701	ATP1A1	ATPase, Na+K+ transporting, alpha 1
'-'			' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' '	polypeptide
428	220973_s_at	NM 030974		hypothetical protein DKFZp434N1923
429	220974_x_at	NM 030971	BA108L7.2	similar to rat tricarboxylate carrier-like
430	220980_s_at	NM 031284		hypothetical protein DKFZp434B195
431	221059 s at	NM 021615	CHST6	carbohydrate sulfotransferase 6
432	221483 s at	AF084555	ARPP-19	okadaic acid-inducible and cAMP-
102	221100_0_u	7 00 1000	1	regulated phosphoprotein 19
433	221484 at	NM 004776		UDP-Gal:betaGlcNAc beta 1,4-
	a.	''''_		galactosyltransferase, polypeptide 5
434	221538_s_at	AL136663		DKFZp564A176
435	221558 s at	AF288571	LEF1	lymphoid enhancer factor-1
436	221577_x_at	AF003934		prostate differentiation factor
437	221641 s at	AF241787		CGI16-iso
438	221688 s at	AL136913		DKFZp586L0118
439	221710 x at	BC006241		hypothetical protein FLJ10647
440	221732_at	AK026161		FLJ22508 fis
441	221759 at	AL583123		CLONE=CS0DL009YN09
442	221797 at	AY007126		clone CDABP0028
443	221799 at	AB037823		KIAA1402 protein,
444	221815 at	BE671816		ESTs
445	221882 s at	Al636233		five-span transmembrane protein M83
446	221902_o_ut	AL567940		CLONE=CS0DF036YK19
447	221909_at	BF984207		ESTs
448	221962_s_at	Al829920		ubiquitin-conjugating enzyme E2H
449	222116_s_at	AL157485		DKFZp762O207
450	222110 s at	AK023133		FLJ13071 fis
451	222155 s at	AK021918		FLJ11856 fis
452	222175 s at	AK000003		FLJ00003 protein
453	222175 s at	AK000470		FLJ20463 fis
454	222199_s_at	AK000470 AK001289		FLJ10427 fis
455	222206_s_at	AA781143		EUROIMAGE 2021883
456	222212_s_at	AK001105		FLJ10243 fis
457		AK001105 AK025328		FLJ10243 its FLJ21675 fis
457	222231_s_at			
	222234 s at	AK022644 AL137749		FLJ12582 fis
459	222240_s_at	AL 13//49		DKFZp434A0612

400	000004 = -/	LAW074445		TEGE
460	222294 s_at	AW971415	 	ESTs
461	32811_at	X98507	TVDO	myosin-I beta
462	40560_at	U28049	TXB2	TBX2
463	44783 s at	R61374		IMAGE-37665
464	46665_at	Al949392		IMAGE-2470926
465	55093_at	AA534198		IMAGE-993116
466	_63825_at	Al557319		
467	87100_at	Al832249		
468	200017_at	NM_002954	RPS27A	ribosomal protein S27a
469	200606_at	NM_004415	DSP	desmoplakin (DPI, DPII)
470	200632 s at	NM_006096	NDRG1	N-myc downstream regulated
471	200636_s_at	NM_002840	PTPRF	protein tyrosine phosphatase,
				receptor type, F
472	200795_at	NM_004684	SPARCL1	SPARC-like 1
473	200810_s_at	NM_001280	CIRBP	cold inducible RNA-binding protein
474	200897_s_at	NM_016081	KIAA0992	Palladin
475	200953_s_at	NM_001759	CCND2	cyclin D2
476	200965_s_at	NM_006720	ABLIM-s	actin binding LIM protein 1 transcript
				variant
477	201012_at	NM_000700	ANXA1	annexin A1
478	201041_s_at	NM_004417	DUSP1	dual specificity phosphatase 1
479	201125_s_at	NM_002213	ITGB5	integrin, beta 5
480	201200_at	NM_003851	CREG	cellular repressor of E1A-stimulated
				genes
481	201286_at	Z48199	syndecan 1	syndecan-1 gene (exons 2-5)
482	201328_at	AL575509		v-ets avian erythroblastosis virus E26
	·			oncogene homolog 2
483	201425_at	NM_000690	ALDH2	aldehyde dehydrogenase 2,
				mitochondrial
484	201427 s_at	NM_005410	SEPP1	selenoprotein P, plasma, 1
485	201432_at	NM_001752	CAT	Catalase
486	201540_at	NM_001449	FHL1	four and a half LIM domains 1
487	201667_at	NM_000165	GJA1	gap junction protein, alpha 1, 43kD
488	201681 s at	AB011155	KIAA0583	KIAA0583
489	201798 s_at	NM_013451	FER1L3	fer-1 (C.elegans)-like 3 (myoferlin)
490	201820_at	NM_000424	KRT5	keratin 5
491	201829_at	AW263232	NET1	neuroepithelial cell transf gene 1
492	201830 s_at	NM_005863	NET1_	neuroepithelial cell transf gene 1
493	201839_s_at	NM_002354	TACSTD1	tumor-associated calcium signal
				transducer 1
494	201842_s_at	Al826799		EGF-containing fibulin-like
				extracellular matrix protein 1
495	201843_s_at	NM_004105	EFEMP1	EGF-containing fibulin-like
				extracellular matrix protein 1
Ĺ				transcript variant 1
496	201983_s_at	AW157070		epidermal growth factor receptor
497	201984 s at	NM_005228	EGFR	epidermal growth factor receptor
498	202054_s_at	NM_000382	ALDH3A2	aldehyde dehydrogenase 3 family,
				member A2
499	202085_at	NM_004817	TJP2	tight junction protein 2
500	202193_s_at	NM_001144	AMFR	autocrine motility factor receptor
501	202196_s_at	NM_013253	DKK3	dickkopf (Xenopus laevis) homolog 3
502	202242_at	NM_004615	TM4SF2	transmembrane 4 superfamily mem 2
503	202267_at	NM_005562	LAMC2	laminin, gamma 2, transcript variant 1
<u> 503</u>	202267_at	NM_005562	LAMC2	laminin, gamma 2, transcript variant 1

WO 2006/002433 PCT/US2005/022846

504	202286 s at	J04152	GA733-1	gastrointestinal tumor-assoc antigen
505	202289 s at	NM_006997	TACC2	transforming, acidic coiled-coil
000	a	14111_000007		containing protein 2
506	202350 s at	NM 002380	MATN2	matrilin 2 precursor, transcript var 1
507	202387 at	NM 004323	BAG1	BCL2-associated athanogene
508	202489 s_at	BC005238	10.401	FXYD domain-containing ion
000		00000200		transport regulator 3
509	202525 at	NM 002773	PRSS8	protease, serine, 8 (prostasin)
510	202552 s at	NM 016441	CRIM1	cysteine-rich motor neuron 1
511	202565 s at	NM 003174	SVIL	supervillin transcript variant 1
512	202575 at	NM 001878	CRABP2	cellular retinoic acid-binding protein 2
513	202597 at	AU144284		interferon regulatory factor 6
514	202668 at	BF001670		ephrin-B2
515	202712 s at	NM 020990	CKMT1	creatine kinase, mitochondrial 1
				nuclear gene mitochondrial protein
516	202746 at	AL021786		PAC 696H22 on chrom Xq21.1-21.2
517	202826 at	NM 003710	SPINT1	serine protease inhibitor, Kunitz t 1
518	202890_at	T62571		microtubule-associated protein 7
519	202936_s_at	NM_000346	SOX9	SRY-box 9
520	202994 s at	Z95331		clone CTA-941F9 on chrom 22q13
521	203037_s_at	NM_014751	KIAA0429	KIAA0429 gene product
522	203074_at	NM_001630	ANXA8	annexin A8
523	203081_at	NM_020248	LOC56998	beta-catenin-interacting protein ICAT
524	203126_at	NM_014214	IMPA2	inositol(myo)-1(or 4)-
				monophosphatase 2
525	203178_at	NM_001482	GATM	glycine amidinotransferase
526	203240_at	NM_003890	FC(γ)BP	IgG Fc binding protein
527	203327_at	N22903		insulin-degrading enzyme
528	203355_s_at	NM_015310	KIAA0942	KIAA0942 protein
529	203407_at	NM_002705	PPL	Periplakin
530	203408_s_at	NM_002971	SATB1	special AT-rich sequence binding protein 1
531	203430_at	NM_014320	SOUL	putative heme-binding protein
532	203453_at	NM_001038	SCNN1A	Na channel, nonvoltage-gated 1 α
533	203485_at	NM_021136	RTN1	reticulon 1
534	203549_s_at	NM_000237	LPL	lipoprotein lipase
535	203571_s_at	NM_006829	APM2	adipose specific 2
536	203585_at	NM_007150	ZNF185	zinc finger protein 185 (LIM domain)
537	203636_at	BE967532	MID1	midline 1 (OpitzBBB syndrome)
538	203637_s_at	NM_000381	MID1	
539	203638_s_at	NM_022969	FGFR2	FGF receptor 2 transcript var 2
540	203678_at	NM_014967	KIAA1018	KIAA1018 protein
541	203687_at	NM_002996	SCYD1	small inducible cytokine subfam D (Cys-X3-Cys) mem 1
542	203726_s_at	NM_000227	LAMA3	laminin, alpha 3
543	203786_s_at	NM_003287	TPD52L1	tumor protein D52-like 1
544	203797_at	AF039555	VSNL1	visinin-like protein 1
545	203799_at	NM_014880	KIAA0022	KIAA0022 gene product
546	203812_at	AB011538		MEGF5
547	203881_s_at	NM_004010		dystrophin transcript variant Dp427p2
548	203910_at	NM_004815	PARG1	PTPL1-associated RhoGAP 1
549	203917_at	NM_001338	CXADR	coxsackie virus and adenovirus receptor
550	203961 at	AL157398	NEBL	nebulette protein
200	1 20000 1 at	AL 107000	(14PAP	Hongierre hioreiti

554	0000001	L NIM	NEOL	
551	203962_s_at	NM_006393	NEBL	<u> </u>
552	203963_at	NM_001218	CA12	carbonic anhydrase XII
553	203992_s_at	AF000992	UTX	ubiquitous TPR motif, X isoform
554	000007	1114 000000	DTDNIC	alternative transcript 1
554	203997_at	NM_002829	PTPN3	protein tyrosine phosphatase, non-
				receptor type 3
555	204005_s_at	NM 002583	PAWR	PRKC, apoptosis, WT1, regulator
556	204019_s_at	NM 015677		hypothetical protein DKFZP586F1318
557	204036_at	AW269335	ĺ	endothelial differentiation,
				lysophosphatidic acid G-protein-
				coupled receptor, 2
558	204037_at	AW269335		endothelial differentiation,
İ			İ	lysophosphatidic acid G-protein-
				coupled receptor, 2
559	204042_at	AB020707		KIAA0900 protein
560	204058_at	AL049699		clone 747H23 on chrom 6q13-15
561	204059_s_at	NM_002395	ME1	malic enzyme 1, NADP(+)-dependent,
				cytosolic
562	204072_s_at	NM_023037	13CDNA73	putative gene product
563	204112_s_at	NM_006895	HNMT	histamine N-methyltransferase
564	204135_at	NM_014890	DOC1	downregulated in ovarian cancer 1
565	204136_at	NM_000094	COL7A1	collagen, type VII, alpha 1
566	204151_x_at	NM_001353	AKR1C1	aldo-keto reductase family 1, mem C1
567	204154_at	NM_001801	CDO1	cysteine dioxygenase, type l
568	204168_at	NM_002413	MGST2	microsomal glutathione S-transferase 2
569	204201 s at	NM 006264	PTPN13	protein tyrosine phosphatase, non-
		_		receptor type 13
570	204204_at	NM_001860	SLC31A2	solute carrier family 31 member 2
571	204224_s_at	NM_000161	GCH1	GTP cyclohydrolase 1
572	204254_s_at	NM_000376	VDR	vitamin D receptor
573	204345_at	NM_001856	COL16A1	collagen, type XVI, alpha 1
574	204351_at	NM_005980	S100P	S100 calcium-binding protein P
575	204359_at	NM_013231	FLRT2	fibronectin leucine rich
		_		transmembrane protein 2
576	204363_at	NM_001993	F3	coagulation factor III
577	204379 s_at	NM_000142	FGFR3	fibroblast growth factor receptor 3
578	204388_s_at	NM_000240	MAOA	monoamine oxidase A
579	204389_at	NM_000240	MAOA	monoamine oxidase A
580	204400_at	NM_005864	EFS2	signal transduction protein
581	204421_s_at	M27968	FGF	basic fibroblast growth factor
582	204422 s_at	NM_002006	FGF2	fibroblast growth factor 2 (basic)
583	204424_s_at	AL050152		neuronal specific transcription factor
				DAT1
584	204455_at	NM_001723	BPAG1	bullous pemphigold antigen 1
585	204503 at	NM 001988	EVPL	envoplakin
586	204517_at	BE962749	cyclophilin C	peptidylprolyl isomerase C
587	204519_s_at	NM_015993	LOC51090	plasmolipin
588	204537_s_at	NM 004961	GABRE	gamma-aminobutyric acid A receptor,
				epsilon transcript variant 1
589	204591_at	NM 006614	CHL1	cell adhesion molecule with homology
•				to L1CAM
590	204600_at	NM 004443	EPHB3	EphB3
591	204671 s at	BE677131		KIAA0957 protein
592	204675 at	NM_001047	SRD5A1	steroid-5-a-reductase, a polypeptide 1
<u> </u>	at	14141 00 10-11	I CINDON I	Totoroid ord-roddoidae, a polypeptide T

593	204718 at	NM 004445	EPHB6	L-PB6
593	204718_at	NM 007168	ABCA8	EphB6 ATP-binding cassette, sub-fam A
394	2047 13_at	14101_007 108	ABCAO	mem 8
595	204734 at	NM 002275	KRT15	keratin 15
596	204749 at	NM 004538	NAP1L3	nucleosome assembly protein 1-like 3
597	204753 s at	Al810712	IVAL ILS	hepatic leukemia factor
598	204753 s_at	Al810712		hepatic leukemia factor
599	204755 x at	M95585	HLF	leukemia factor
600	204765_x_at	NM_005435	ARHGEF5	Rho guanine nucleotide exchange
000	204705_at	14141_003433	AKIIGEI 3	factor 5
601	204773 at	NM 004512	IL11RA	interleukin 11 receptor, alpha
602	204773 at	NM 004512	IL11RA	
603	204855_at	NM 002639	SERPINB5	serine (or cysteine) proteinase
		l		inhibitor, clade B, member 5
604	204872_at	NM_007005	BCE-1	BCE-1 protein
605	204937_s_at	NM_016325	ZNF274	zinc finger protein 274
606	204942_s_at	NM_000695	ALDH3B2	aldehyde dehydrogenase 3 fam mem
				B2
607	204952_at	NM_014400	C4.4A	GPI-anchored metastasis-associated
				protein homolog
608	204971_at	NM_005213	CSTA	cystatin A (stefin A)
609	204975_at	NM_001424	EMP2	epithelial membrane protein 2
610	204990_s_at	NM_000213	ITGB4	integrin, beta 4
611	205014_at	NM_005130	HBP17	heparin-binding growth factor binding
612	205019_s_at	NM_004624	VIPR1	vasoactive intestinal pep receptor 1
613	205081_at	NM_001311	CRIP1	cysteine-rich protein 1 (intestinal)
614	205109_s_at	NM_015320	ARHGEF4	Rho guanine nucleotide exchange
615	205128 x at	NM_000962	PTGS1	factor (GEF) 4 prostaglandin-endoperoxide synthase 1
616	205120_X_at	NM_006846	SPINK5	serine protease inhibitor, Kazal t, 5
617	205105_at	NM 003278	TNA	tetranectin
618	205206 at	NM_000216	KAL1	Kallmann syndrome 1 sequence
619	205236 x at	NM_003102	SOD3	superoxide dismutase 3, extracellular
620	205251_at	NM_022817	PER2	period homolog 2 transcript variant 1
621	205259 at	NM 000901	NR3C2	nuclear receptor subfamily 3, group
021	200200_at	11111_000001	MICOZ	C, member 2
622	205286 at	U85658		transcription factor ERF-1
623	205349_at	NM 002068	GNA15	guanine nucleotide binding protein, α 15
624	205363 at	NM 003986	BBOX1	butyrobetaine (γ), 2-oxoglutarate
				dioxygenase 1
625	205382_s_at	NM_001928	DF	D component of complement (adipsin)
626	205384_at	NM_005031	FXYD1	FXYD domain-containing ion
	<u> </u>		<u> </u>	transport regulator 1 variant a
627	205403_at	NM_004633	IL1R2	interleukin 1 receptor, type II
628	205404_at	NM_005525	HSD11B1	hydroxysteroid dehydrogenase 1
629	205407_at	NM_021111	RECK	reversion-inducing-cysteine-rich
				protein with kazal motifs
630	205440_s_at	NM_000909	NPY1R	neuropeptide Y receptor Y1
631	205455_at	NM_002447	MST1R	macrophage stimulating 1 receptor
632	205464_at	NM_000336	SCNN1B	Na channel, nonvoltage-gated 1, β
633	205470_s_at	NM_006853	KLK11	kallikrein 11
634	205490_x_at	BF060667	connexin 31	gap junction protein, beta 3, 31kD
635	205498_at	NM_000163	GHR	growth hormone receptor
636	205559 s at	NM_006200	PCSK5	proprotein convertase subtilisinkexin

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		<u> </u>		type 5
637	205560_at	NM_006200	PCSK5	
638	205569_at	NM_014398	TSC403	similar to lysosome-associated
				membrane glycoprotein
639	205613_at	NM_016524	LOC51760	BK protein
640	205668_at	NM_002349	LY75	lymphocyte antigen 75
641	205672_at	NM_000380	XPA	xeroderma pigmentosum,
				complementation group A
642	205709_s_at	NM_001263_	CDS1	CDP-diacylglycerol synthase 1
643	205730_s_at	NM_014945	KIAA0843	KIAA0843 protein
644	_205765_at	NM_000777	CYP3A5	cyt P450, subfam IIIA, polypep 5
645	205807_s_at	NM_020127	TUFT1	tuftelin 1
646	205857_at	Al269290		solute carrier family 18, member 2
647	205883_at	NM_006006	ZNF145	zinc finger protein 145
648	205900_at	NM 006121	KRT1	keratin 1
649	205933_at	NM 015559	KIAA0437	KIAA0437 protein
650	205977 s at	NM 005232	EPHA1	EphA1
651	206032 at	Al797281		est:we86g02.x1
652	206033_s_at	NM 001941	DSC3	desmocollin 3 transcript variant Dsc3a
653	206068_s_at	Al367275		enzyme A dehydrogenase, long chain
654	206093_x_at	NM 007116	TNXA	tenascin XA
655	206122_at	NM_006942	SOX20	SRY-box 20
656	206149 at	NM_022097	LOC63928	hepatocellular carcinoma antigen
				gene 520
657	206170 at	NM 000024	ADRB2	adrenergic, beta-2-, receptor, surface
658	206192 at	L20815		S protein
659	206201 s at	NM_005924	MEOX2	mesenchyme homeo box 2
660	206276_at	NM_003695	E48	lymphocyte antigen 6 comp locus D
661	206315 at	NM_004750	CRLF1	cytokine receptor-like factor 1
662	206363_at	NM 005360	MAF	v-maf musculoaponeurotic
				fibrosarcoma oncogene homolog
663	206385 s at	NM_020987	ANK3	ankyrin 3, node of Ranvier, tran var 1
664	206400_at	NM 002307	LGALS7	lectin, galactoside-binding, soluble, 7
665	206453_s_at	NM_016250	NDRG2	N-myc downstream-regulated gene 2
666	206481_s_at	NM_001290	LDB2	LIM domain binding 2
667	206482_at	NM_005975	PTK6	PTK6 protein tyrosine kinase 6
668	206515_at	NM_000896	CYP4F3	cyt P450, subfam IVF, polypeptide 3
669	206539 s at	NM 023944	CYP4F12	cytochrome P450 isoform 4F12
670	206581_at	NM 001717	BNC	basonuclin
671	206637_at	NM_014879	KIAA0001	KIAA0001 gene product
672	206655 s at	NM 000407	GP1BB	glycoprotein lb (platelet), ß polypep
673	206693_at	NM_000880	IL7	interleukin 7
674	206884 s at	NM 003843	SCEL.	sciellin
675	207002_s_at	NM 002656	PLAGL1	pleiomorphic adenoma gene-like 1
676	207002_s_at	NM_000421	KRT10	keratin 10
677	207076_s_at	NM 000050	ASS	argininosuccinate synthetase
678	207076 s at	NM 002748	MAPK6	mitogen-activated protein kinase 6
679	207655_s_at	NM_013314	SLP65	B cell linker protein
680	207720_at	NM_000427	LOR	loricrin
681	207761 s_at	NM_014033	0)/05	DKFZP586A0522 protein
682	207843 x at	NM_001914	CYB5	cytochrome b-5
683	207908_at	NM_000423	KRT2A	keratin 2A
684	207943_x_at	NM_006718	PLAGL1	pleiomorphic adenoma gene-like
	<u></u>			transcript variant 2

685	207955_at	NM_006664	SCYA27	small inducible cytokine subfamily A
				(Cys-Cys), member 27
686	207996_s_at	NM_004338	C18ORF1	chrom 18 open reading frame 1
687	208096 s at	NM_030820		hypothetical protein DKFZp564B052
688	208146_s_at	NM_031311	LOC54504	serine carboxypeptidase vitellogenic- like
689	208161_s_at	NM 020037	ABCC3	ATP-binding cassette sub-fam C mem 3
690	208190_s_at	NM 015925	LISCH7	liver-specific bHLH-Zip transcription
L				factor
691	208228 s_at	M87771	K-sam-III	secreted FGF receptor
692	208609_s_at	NM_019105	TNXB	tenascin XB
693	208614_s_at	M62994		thyroid autoantigen
694	208651_x_at	M58664		CD24 signal transducer
695	208690 s at	BC000915		Similar to LIM protein,
696	208798_x_at	AF204231	GM88	88-kDa Golgi protein
697	209047_at	AL518391		aquaporin 1
698	209159_s_at	AV724216		NDRG family, member 4
699	209160_at	AB018580	hluPGFS	aldo-keto reductase family 1, mem C3
700	209211_at	AF132818	CKLF	colon Kruppel-like factor
701	209212_s_at	AB030824		transcription factor BTEB2
702	209289_at	Al700518		nuclear factor IB
703	209290_s_at	BC001283		Similar to nuclear factor IB,
704	209309_at	D90427		zinc-alpha2-glycoprotein
705	209318_x_at	BG547855		pleiomorphic adenoma gene-like 1
706	209335_at	Al281593	-	decorin
707	209348_s_at	AF055376	c-maf	short form transcription factor C-MAF
708	209351_at	BC002690		keratin 14
709	209357_at	AF109161	MRG1	p35srj
710	209366_x_at	M22865		cytochrome b5
711	209368_at	AF233336	EPHX2	soluble epoxide hydrolase
712	209386_at	Al346835		transmembrane 4 superfam mem 1
713	209392_at	L35594	autotaxin	ectonucleotide
				pyrophosphatasephosphodiesterase 2
714	209465_x_at	AL565812		pleiotrophin
715	209493_at	AF338650	AIPC	PDZ domain-containing protein AIPC
716	209540_at	NM_000618	somatomedin (
717	209550_at	U35139		NECDIN related protein
718	209558_s_at	AB013384	HIP1R	huntingtin interacting protein-1-related
719	209590_at	AL157414		clone RP11-560A15 on chrom 20
720	209602_s_at	AI796169		GATA-binding protein 3
721	209603_at	Al796169		GATA-binding protein 3
722	209604_s_at	BC003070		GATA-binding protein 3, clone MGC:2346
723	209605 at	D87292	rhodanese	thiosulfate sulfurtransferase
724	209656_s_at	AL136550		DKFZp761J17121
725	209679_s_at	BC003379	_	hyp protein from clone 643, clone MGC:5115
726	209684 at	AL136924		DKFZp586G2120
727	209687 at	U19495	hIRH	intercrine-alpha
728	209691_s_at	BC003541	FLJ10488	hypothetical protein FLJ10488
729	209699_x_at	U05598		dihydrodiol dehydrogenase
730	209732 at	NM 005127	 	Sim to C-type lectin, superfam mem 2
731	209763 at	AL049176		clone 141H5 on chrom Xq22.1-23
732	209771_x_at	AA761181	 	CD24 antigen
132	ZUSIII X at	MA/01101	l	L CDZ4 antigen

733	209863 s at	AF091627	T	CUSP
734	209866 s_at	R50822		KIAA0768 protein
735	209975 at	AF182276	CYP2E1	cytochrome P450-2E1
736	210059_s_at	BC000433	1011.221	mitogen-activated protein kinase 13
737	210096_at	J02871	 	lung cytochrome P450 Bl
738	210128 s at	U41070		P2 purinergic receptor
739	210298 x at	AF098518	FHL1	4 and ½ LIM domains 1 protein isoform
733	210230_x_at	71 030010	1116	B
740	210347_s_at	AF080216		C2H2-type zinc-finger protein
741	210377 s at	AF208012	TPD52L2	tumor protein D52-like 2
742	210397 at	U73945	11 00222	beta-defensin-1
743	210619_s_at	AF173154	HYAL1	hyaluronoglucosaminidase 1 isof 2
744	210633_x_at	M19156	KRT10	acidic keratin-10
745	210715 s at	AF027205	kop	Kunitz-type protease inhibitor
746	210880 s at	AB001467	Efs2	Numez type protease in libitor
747	210958 s at	BC003646	LIGE	clone MGC:4693
748	211043 s at	BC006332	Lcb	clathrin, light polypeptide
749	2111045_s_at	U80918	NF-ATcC	transcription factor
750	211382 s at	AF220152	TACC2	transforming, acidic coiled-coil
' 30	211002_3_at	74 220102	17.002	containing protein 2
751	211458 s at	AF180519		GABA-A receptor-associated
752	211596 s at	AB050468		membrane glycoprotein LIG-1
753	211597 s at	AB059408	SMAP31-12	
754	211653 x at	M33376		pseudo-chlordecone reductase
755	211712_s_at	BC005830		clone MGC:1925
756	211734 s at	BC005912	 	Fc fragment IgE, high affinity I, rec for α
1				polypep
757	211841 s at	U94510		lymphocyte associated receptor of
	l			death 9, alternatively spliced
758	211986_at	BG287862	desmoyokin	AHNAK nucleoprotein
759	212148_at	BF967998		FLJ12900 fis,
760	212204_at	AF132733		DKFZP564G2022 protein
761	212242_at	AL565074		tubulin, alpha 1 (testis specific)
762	212327_at	AK027231	ļ	FLJ23578 fis, KIAA1102 protein
763	212328_at	AK027231		FLJ23578 fis, KIAA1102 protein
764	212390_at	AB007923		KIAA0477 gene product
765	212538_at	AL576253		KIAA1058 protein
766	212543_at	U83115.1		non-lens β gamma-crystallin like
767	212589_at	BG168858		oncogene TC21
768	212593_s_at	N92498		FLJ22071 fis, clone HEP11691
769	212724 at	BG054844	<u> </u>	ras homolog gene family, member E
770	212741_at	AA923354		monoamine oxidase A
771	212823_s_at	AU147160		KIAA0599 protein
772	212841_s_at	Al692180		PTPRF interacting protein, binding
775	040050	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	 	protein 2
773	212850_s_at	AA584297		low density lipoprotein receptor-related
774	040075	AD004745	 	protein 4
774	212875_s_at	AP001745	 	chrom 21 open reading frame 25
775	212992_at	Al935123		ESTS
776	213029_at	AL110126		DKFZp564H1916
777	213032_at	AL110126	 	DKFZp564H1916
778	213050_at	AA594937	 	KIAA0633 protein
779	213068_at	A1146848	-	dermatopontin
780	213071_at	AI146848	J	dermatopontin

781	213106_at	Al769688		23664 and 23905 mRNA sequence
782	213110 s_at	AW052179		collagen, type IV, alpha 5
783	213110_s_at	AI096375	 	KIAA1750 protein, partial cds
784	213135 at	U90902		clone 23612
785	213194_at	BF059159		Hs.301198 roundabout homolog
786	213227_at	BE879873	 	progesterone membrane binding
787	213280 at	AK000478		FLJ20471 fis
788	213285 at	AV691491		DKFZp564D1462
789	213287_s_at	X14487	 	acidic (type I) cytokeratin 10
790	213267_s_at	BF693921		ATP-binding cassette, sub-family A,
190	213333_at	05093921		member 5
791	213359_at	W74620		heterogeneous nuclear
' '				ribonucleoprotein D
792	213369 at	Al825832		DKFZp434A132
793	213375 s at	N80918		Novel gene mapping to chomo 13
794	213397 x at	Al761728		DnaJ homolog, subfam C, mem 8
795	213451_x_at	BE044614		tenascin XB
796	213456_at	Al927000		DKFZP564D206
797	213506_at	BE965369		proteinase activated receptor-2
798	213556 at	BE673445		chromosome 19, cosmid R28379
799	213618 at	AB011152		KIAA0580
800	213695 at	L48516	PON3	paraoxonase 3
801	213707_s_at	NM 005221	DLX5	distal-less homeo box 5
802	213725_x_at	Al693140		DKFZp586F071
803	213737 x at	Al620911		
804	213800 at	X04697		complement factor H 38-kDa N-term
805	213817_at	AL049435	•	DKFZp586B0220
806	213820 s at	T54159		hypothetical protein MGC10327
807	213844_at	NM_019102	HOXA5	homeo box A5
808	213848_at	Al655015		DKFZp586F2224
809	213891_s_at	Al927067		FLJ11918 fis
810	213924_at	BF476502		hypothetical protein FLJ11585
811	213929_at	AL050204		DKFZp586F1223
812	213933_at	AW242315		DKFZp586M0723
813	213935_at	AF007132		clone 23551
814	213942_at	AL134303		DKFZp547K034_r1
815	213992_at	Al889941		collagen, type IV, alpha 6
816	213994_s_at	Al885290		spondin 1, extracellular matrix
817	214058_at	M19720		L-myc protein
818	214132_at	BG232034		ATP synthase, H+ transporting,
ŀ				mitochondrial F1 complex, gamma
•	<u> </u>			polypeptide 1
819	214164_x_at	BF752277]	adaptor-related protein complex 1,
			ļ	gamma 1 subunit
820	214234_s_at	X90579		cyp related pseudogene
821	214235_at	X90579		cyp related pseudogene
822	214247 s at	AU148057		regulated in glioma
823	214598_at	AL049977		DKFZp564C122
824	214696_at	AF070569	 	clone 24659
825	214753_at	AW084068	ļ	BRCA2 region
826	214823_at	AF033199		C2H2 zinc finger protein pseudogene
827	215034_s_at	Al189753		FLJ13302 fis
828	215062_at	AL390143		DKFZp547N074 C2 domain containing Pl3-kinase
829	215129 at	AJ000008		

	1045000	1.11100700		1
830	215239_x_at	AU132789	-	zinc finger protein 273
831	215243 s at	AF099730	GJB3	connexin 31
832	215388 s at	X56210	FHR-1	complement Factor H-related 1
833	215513_at	AF241534	HYMAI	hydatidiform mole assoc & imprinted
834	215516_at	AC005048		BAC clone CTB-15P3 fr 7q22-q31.2
835	215536_at			DMA, DMB, HLA-Z1, IPP2, LMP2,
				TAP1, LMP7, TAP2, DOB, DQB2 and
				RING8, 9, 13 and 14 genes
836	215659_at	AK025174		FLJ21521
837	215704_at	AL356504		clone RP1-14N1 chrom 1q21.1-21.3
838	215726_s_at	M22976		cytochrome b5
839	215867_x_at	AL050025		DKFZp564D066
840	216199_s_at	AL109942		clone RP3-473J16 chrom 6q25.3-26
841	216268 s at	U77914		soluble protein Jagged
842	216333_x_at	M25813		unidentified gene complementary to
				P450c21
843	216379_x_at	AK000168		CD24 signal transducer
844	216594_x_at	S68290		chlordecone reductase homolog
845	216699 s at	L10038		pre-pro-protein for kallikrein
846	217087_at	AF005081	хр32	skin-specific protein
847	217528_at	BF003134		ESTs
848	217707_x_at	AI535683		ESTs
849	217901_at	BF031829		desmoglein 2
850	217961_at	NM_017875	FLJ20551	hypothetical protein FLJ20551
851	218002_s_at	NM_004887	SCYB14	small inducible cytokine subfamily B
				(Cys-X-Cys), member 14
852	218170_at	NM_016048	LOC51015	CGI-111 protein
853	218180_s_at	NM_022772	FLJ21935	hypothetical protein FLJ21935
854	218186_at	NM_020387	CATX-8	CATX-8 protein
855	218237_s_at	NM_030674	ATA1	amino acid transporter system A1
856	218326_s_at	NM_018490	GPR48	G protein-coupled receptor 48
857	218434_s_at	NM_023928		hypothetical protein FLJ12389
858	218451_at	NM_022842		hypothetical protein FLJ22969
859	218499_at	NM_016542	LOC51765	serinethreonine protein kinase MASK
860	218546 at	NM_024709		hypothetical protein FLJ14146
861	218552_at	NM_018281		hypothetical protein FLJ10948
862	218603_at	NM 016217	LOC51696	hHDC for homolog of Dros headcase
863	218644 at	NM 016445	PLEK2	pleckstrin 2 (mouse) homolog
864	218651_s_at	NM_018357		hypothetical protein FLJ11196
865	218657_at	NM_016339	LOC51195	Link guanine nucleotide exchange
	_	_		factor II
866	218675_at	NM_020372	LOC57100	organic cation transporter
867	218677_at	NM_020672	LOC57402	S100-type calcium binding protein A14
868	218692_at	NM_017786		hypothetical protein FLJ20366
869	218704_at	NM 017763		hypothetical protein FLJ20315
870	218718 at	NM_016205	PDGFC	platelet derived growth factor C
871	218736_s_at	NM 017734		hypothetical protein FLJ20271
872	218751_s_at	NM_018315		hypothetical protein FLJ11071
873	218764_at	NM_024064		hypothetical protein MGC5363
874	218792 s at	NM 017688		hypothetical protein FLJ20150
875	218796_at	NM_017671		hypothetical protein FLJ20116
876	218804 at	NM 018043		hypothetical protein FLJ10261
877	218806_s_at	AF118887	VAV-3	VAV-3 protein
878	218807 at	NM 006113	VAV3	vav 3 oncogene
<u> </u>	1			1 3 0,100,901.0

	1040040	T	т	T
879	218816_at	NM_018214		hypothetical protein FLJ10775
880	218820_at	NM_020215		hypothetical protein DKFZp761F2014
881	218849 s_at	NM_006663	RAI	RelA-associated inhibitor
882	218854_at	NM_013352	SART-2	squamous cell carcinoma antigen recog
				by T cell
883	218901_at	NM_020353	LOC57088	phospholipid scramblase 4
884	218919_at	NM_024699		hypothetical protein FLJ14007
885	218963_s_at	NM_015515		DKFZP434G032 protein
886	219010_at	NM_018265		hypothetical protein FLJ10901
887	219054_at	NM_024563		hypothetical protein FLJ14054
888	219064_at	NM_030569		hypothetical protein MGC10848
889	219073 s_at	NM_017784		hypothetical protein FLJ20363
890	219090_at	NM_020689	NCKX3	sodium calcium exchanger
891	219093_at	NM_017933		hypothetical protein FLJ20701
892	219095_at	NM_005090	PLA2G4B	phospholipase A2, group IVB
893	219109_at	NM_024532		hypothetical protein FLJ22724
894	219115_s_at	NM_014432	IL20RA	interleukin 20 receptor, alpha
895	219229_at	NM_013272	SLC21A11	solute carrier family 21, member 11
896	219232_s_at	NM_022073	ļ	hypothetical protein FLJ21620
897	219263_at	NM_024539		hypothetical protein FLJ23516
898	219298_at	NM_024693		hypothetical protein FLJ20909
899	219313_at	NM_017577		hypothetical protein DKFZp434C0328
900	219368_at	NM_021963	NAP1L2	nucleosome assembly protein 1-like 2
901	219388_at	NM_024915		hypothetical protein FLJ13782
902	219395_at	NM_024939		hypothetical protein FLJ21918
903	219410_at	NM_018004		hypothetical protein FLJ10134
904	219411_at	NM_024712		hypothetical protein FLJ13824
905	219423_x_at	NM_003790	TNFRSF12	TNF receptor superfamily, member 12
906	219436_s_at	NM_016242	LOC51705	endomucin-2
907	219461_at	AJ236915		pak5 protein
908	219476_at	NM_024115		hypothetical protein MGC4309
909	219489 s_at	NM_017821		hypothetical protein FLJ20435
910	219497 s at	NM_022893	BCL11A	B-cell CLLlymphoma 11A
911	219518_s_at	NM_025165		hypothetical protein FLJ22637
912	219528_s_at	NM_022898	BCL11B	B-cell lymphomaleukaemia 11B
913	219532_at	NM_022726	ELOVL4	Stargardt disease 3
914	219597_s_at	NM_017434	DUOX1	dual oxidase 1
915	219689_at	NM_020163	LOC56920	semaphorin sem2
916	219729_at	NM_016307	PRX2	paired related homeobox protein
917	219764_at	NM_007197	FZD10	frizzled (Drosophila) homolog 10
918	219806 s_at	NM_020179	FN5	FN5 protein
919	219825_at	NM_019885	P450RAI-2	cyt P450 retinoid metabolizing
920	219908_at	NM_014421	DKK2	dickkopf homolog 2
921	219936_s_at	NM_023915	GPR87	G protein-coupled receptor 87
922	219938_s_at	NM_024430	PSTPIP2	proline-serine-threonine phosphatase
022	210070 04	NIM 047655	 	interacting protein 2
923	219970_at	NM_017655	HOOKI	hypothetical protein FLJ20075
924	219976_at	NM_015888	ноок1	hook1 protein
925	219995 s_at	NM_024702		hypothetical protein FLJ13841
926	219998 at	NM 014181		HSPC159 protein
927	220016_at	NM_024060	11 22D	hypothetical protein MGC5395
928	220056_at	NM_021258	IL22R	interleukin 22 receptor
929 930	220066_at 220076_at	NM_022162	NOD2	NOD2 protein
	こくさいしょう おも	NM_019847	ANKH	ankylosis, progressive homolog

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	T 000404	1 224 040444	L = / 13 40	
931	220161 s at	NM_019114	EHM2	EHM2 gene
932	220225_at	NM_016358	IRX4	iroquois homeobox protein 4
933	220230 s at	NM_016229	LOC51700	cytochrome b5 reductase b5R.2
934	220262 s_at	NM_023932		hypothetical protein MGC2487
935	220266 s at	NM_004235	KLF4	Kruppel-like factor 4 (gut)
936	220289 s at	NM_017977		hypothetical protein FLJ10040
937	220318_at	NM_017957	FLJ20778	epsin 3
938	220413_at	NM_014579	ZIP2	zinc transporter
939	220414_at	NM_017422	CLSP	calmodulin-like skin protein
940	220428_at	NM_015717	LANGERIN	Langerhans cell specific c-type lectin
941	220432 s at	NM_016593	CYP39A1	oxysterol 7alpha-hydroxylase
942	220518_at	NM_024801		hypothetical protein FLJ21551
943	220625 s at	AF115403		Ets transcription factor ESE-2b
944	220723 s at	NM_025087		hypothetical protein FLJ21511
945	220724_at	NM_025087		hypothetical protein FLJ21511
946	220911_s_at	NM_025081		KIAA1305 protein
947	220945_x_at	NM_018050		hypothetical protein FLJ10298
948	221127_s_at	NM_006394	RIG	regulated in glioma
949	221215_s_at	NM_020639	ANKRD3	ankyrin repeat domain 3
950	221541_at	AL136861		DKFZp434B044
951	221667_s_at	AF133207		protein kinase H11
952	221747_at	AL046979		DKFZp586K0617
953	221748_s_at	AL046979		DKFZp586K0617
954	221760_at	BG287153		mannosidase, α, class 1A, member 1
955	221796_at	AA707199		Similar to hyp protein FLJ20093
956	221841_s_at	BF514079		Kruppel-like factor 4 (gut)
957	221854_at	Al378979		ESTs
958	221922_at	AW195581		KIAA0761
959	221950_at	Al478455		empty spiracles homolog 2
960	222043_at	Al982754		clusterin
961	222102_at	NM_000847	GSTA3	glutathione S-transferase A3
962	222236_s_at	AK000253		FLJ20246 fis
963	222256 s at	AK000550		FLJ20543 fis
964	222288_at	Al004009		ESTs
965	222290_at	AA731709		ESTs
966	222303_at	AV700891		ESTs
967	266_s_at	L33930		CD24 signal transducer
968	33322_i_at	X57348		clone 9112
969	33323 r at	X57348		clone 9112
970	35666_at	U38276		semaphorin III family homolog
971	38340_at	AB014555	<u> </u>	KIAA0655 protein
972	_39248_at	N74607		za55a01.s1
973	40016_g_at	AB002301		KIAA0303 gene
974	40093_at	X83425		LU gene Lutheran blood group
				glycoprotein
975	40472_at	AF007155		clone 23763 unknown mRNA
976	57588_at	R62432		yg52e11.s1
977	60474_at	AA469071		ne17f11.s1
978	91826_at	Al219073		qg16e08.x1
979			PBGD	
980			MART1	
981			Me20m	
982		ļ	MAGE-3	
983	<u> </u>	Ĺ	L	Me20m forward primer

984	Me20m reverse primer
985	Me20m probe
986	PBGD forward primer
987	PBGD reverse primer
988	PBGD probe
999	Tyrosinase
1000	Tyrosinase Forward
1001	Tyrosinase Reverse
1002	Tyrosinase probe
1003	MART1 Forward
1004	MART1 Reverse
1005	MART1 Probe
1006	HMB45 Forward
1007	gp100 Reverse
1008	gp100 Probe
1009	PLAB Forward
1010	PLAB Reverse
1011	PLAB Probe

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6,475,727 6,500,638 6,521,227 6,527,560 6,599,699

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